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# UTILITY PATENT APPLICATION TRANSMITTAL

(Only for new nonprovisional applications under 37 C.F.R. § 1.53(b))

Attorney Docket No. **NMEDP001-2**  
First Inventor or Application Identifier **Snutch, Terry P.**  
Title **Novel Human Calcium Channels and...**  
Express Mail Label No. **EL362857074US**

## APPLICATION ELEMENTS

See MPEP chapter 600 concerning utility patent application contents

ADDRESS TO: **Box Patent Application  
Washington, DC 20231**

1. ☒ \* Fee Transmittal Form (e.g., PTO/SB/17)  
(Submit an original and a duplicate for fee processing)
2. ☒ Specification [Total Pages **31**]  
(preferred arrangement set forth below)  
- Descriptive title of the Invention  
- Cross References to Related Applications  
- Statement Regarding Fed sponsored R & D  
- Reference to Microfiche Appendix  
- Background of the Invention  
- Brief Summary of the Invention  
- Brief Description of the Drawings (if filed)  
- Detailed Description  
- Claim(s)  
- Abstract of the Disclosure
3. ☒ Drawing(s) (35 U.S.C. 113) [Total Sheets **4**]
4. Oath or Declaration [Total Pages **3**]  
a. ☒ Newly executed (original or copy)  
b. ☐ Copy from a prior application (37 C.F.R. § 1.63(d))  
(for continuation/divisional with Box 16 completed)  
i. ☐ **DELETION OF INVENTOR(S)**  
Signed statement attached deleting  
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see 37 C.F.R. §§ 1.63(d)(2) and 1.33(b).

5. ☐ Microfiche Computer Program (Appendix)
6. Nucleotide and/or Amino Acid Sequence Submission  
(if applicable, all necessary)  
a. ☒ Computer Readable Copy  
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## ACCOMPANYING APPLICATION PARTS

7. ☐ Assignment Papers (cover sheet & document(s))
8. ☐ 37 C.F.R. § 3.73(b) Statement ☒ Power of Attorney  
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10. ☐ Information Disclosure Statement (IDS)/PTO-1449 ☐ Copies of IDS Citations
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Prior application information. Examiner **Basu, N.** Group / Art Unit **1646**

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Oppedahl & Larson

Applicant or Patentee: SNUTCH ET AL Attorney's Docket No. NMED P-001-2  
Serial or Patent No.: TBA Filed or Issued: HEREWITH  
For: NOVEL HUMAN CALCIUM CHANNELS AND RELATED PROBES, CELL LINES AND METHODS

**VERIFIED STATEMENT (DECLARATION) CLAIMING SMALL ENTITY STATUS**  
(37 CFR 1.9(f) and 1.27(e)) - SMALL BUSINESS CONCERN

I hereby declare that I am

- ☐ the owner of the small business concern identified below:  
☒ an official of the small business concern empowered to  
act on behalf of the concern identified below:

NAME OF CONCERN NeuroMed Technologies Inc.  
400 - 601 West Broadway V5Z 4C2  
ADDRESS OF CONCERN 5803 W. 24<sup>th</sup> Avenue, Vancouver, Canada V6S 1M1

I hereby declare that the above identified small business concern qualifies as a small business concern as defined in 13 CFR 121.3-18, and reproduced in 37 CFR 1.9(d), for purposes of paying reduced fees under Section 41(a) and (b) of Title 35, United States Code, in that the number of employees of the concern, including those of its affiliates, does not exceed 500 persons. For purposes of this statement, (1) the number of employees of the business concern is the average over the previous fiscal year of the concern of the persons employed on a full-time, part-time or temporary basis during each of the pay periods of the fiscal year, and (2) concerns are affiliates of each other when either, directly or indirectly, one concern controls or has the power to control the other, or a third party or parties controls or has the power to control both.

I hereby declare that rights under contract or law have been conveyed to and remain with the small business concern identified above with regard to the above-captioned invention which is described in

- ☒ the specification filed herewith  
☐ Application Serial No. \_\_\_\_\_, filed \_\_\_\_\_  
☐ Patent No. \_\_\_\_\_, issued \_\_\_\_\_

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I acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time of paying, the earliest of the issue fee or any maintenance fee due after the date on which status as a small entity is no longer appropriate. (37 CFR 1.28(b))

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NAME OF PERSON SIGNING Natalie Baker TITLE Chief Operating Officer  
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Vancouver, BC V5Z 4C2

SIGNATURE [Signature] DATE 99/07/02

NOVEL HUMAN CALCIUM CHANNELS AND  
RELATED PROBES, CELL LINES AND METHODS

This application is a continuation-in-part of copending US Patent Application No. Serial No. 09/030,482, filed February 25, 1998, which is a 111(a) application claiming priority from US Provisional Application No. 60/039,204, filed February 28, 1997, both of which are incorporated herein by reference.

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**TECHNICAL FIELD**

The present invention relates to novel mammalian (including human) calcium channel compositions, and to the expression of these compositions in cell lines for use in evaluating calcium channel function and the behavior of compositions which modulate calcium channel function.

**BACKGROUND OF THE INVENTION**

The rapid entry of calcium into cells is mediated by a class of proteins called voltage-gated calcium channels. Calcium channels are a heterogeneous class of molecules that respond to depolarization by opening a calcium-selective pore through the plasma membrane. The entry of calcium into cells mediates a wide variety of cellular and physiological responses including excitation-contraction coupling, hormone secretion and gene expression. In neurons, calcium entry directly affects membrane potential and contributes to electrical properties such as excitability, repetitive firing patterns and pacemaker activity. Miller, R.J. (1987) "Multiple calcium channels and neuronal function." *Science* 235:46-52. Calcium entry further affects neuronal functions by directly regulating calcium-dependent ion channels and modulating the activity of calcium-dependent enzymes such as protein kinase C and calmodulin-dependent protein kinase II. An increase in calcium concentration at the presynaptic nerve terminal triggers the release of neurotransmitter. Calcium entry also plays a

role in neurite outgrowth and growth cone migration in developing neurons and has been implicated in long-term changes in neuronal activity.

In addition to the variety of normal physiological functions mediated by calcium channels, they are also implicated in a number of human disorders. Recently, mutations identified in human and mouse calcium channel genes have been found to account for several disorders including, familial hemiplegic migraine, episodic ataxia type 2, cerebellar ataxia, absence epilepsy and seizures. Fletcher, et al. (1996) "Absence epilepsy in tottering mutant mice is associated with calcium channel defects." *Cell* 87:607-617; Burgess, et al. (1997) "Mutation of the Ca<sup>2+</sup> channel  $\beta$  subunit gene Cchb4 is associated with ataxia and seizures in the lethargic (lh) mouse." *Cell* 88:385-392; Ophoff, et al. (1996) "Familial hemiplegic migraine and episodic ataxia type-2 are caused by mutations in the Ca<sup>2+</sup> channel gene CACNL1A4." *Cell* 87:543-552; Zhuchenko, O. et al. (1997) "Autosomal dominant cerebellar ataxia (SCA6) associated with the small polyglutamine expansions in the  $\alpha$ 1A-voltage-dependent calcium channel." *Nature Genetics* 15:62-69.

The clinical treatment of some disorders has been aided by the development of therapeutic calcium channel antagonists. Janis, et al. (1991) in *Calcium Channels: Their Properties, Functions, Regulation and Clinical Relevance*. CRC Press, London.

Native calcium channels have been classified by their electrophysiological and pharmacological properties as T, L, N, P and Q types (for reviews see McCleskey, et al. (1991) "Functional properties of voltage-dependent calcium channels." *Curr. Topics Membr.* 39: 295-326, and Dunlap, et al. (1995) "Exocytotic Ca<sup>2+</sup> channels in mammalian central neurons." *Trends Neurosci.* 18:89-98.). T-type (or low voltage-activated) channels describe a broad class of molecules that activate at negative potentials and are highly sensitive to changes in resting potential. The L, N, P and Q-type channels activate at more positive potentials and display diverse kinetics and voltage-dependent properties. There is some overlap in biophysical properties of the high voltage-activated channels, consequently pharmacological profiles are useful to further distinguish them. L-type channels are sensitive

to dihydropyridine (DHP) agonists and antagonists, N-type channels are blocked by the *Conus geographus* peptide toxin,  $\omega$ -conotoxin GVIA, and P-type channels are blocked by the peptide  $\omega$ -agatoxin IVA from the venom of the funnel web spider, *Agelenopsis aperta*. A fourth type of high voltage-activated Ca channel (Q-type) has been described, although whether the Q- and P-type channels are distinct molecular entities is controversial (Sather et al. (1993)

"Distinctive biophysical and pharmacological properties of class A (B1) calcium channel  $\alpha_1$  subunits." *Neuron* 11: 291-303; Stea, et al. (1994) "Localization and functional properties of a rat brain  $\alpha_1A$  calcium channel reflect similarities to neuronal Q- and P-type channels." *Proc Natl Acad Sci (USA)* 91: 10576-10580.). Several types of calcium conductances do not fall neatly into any of the above categories and there is variability of properties even within a category suggesting that additional calcium channels subtypes remain to be classified.

Biochemical analyses show that neuronal high-threshold calcium channels are heterooligomeric complexes consisting of three distinct subunits ( $\alpha_1$ ,  $\alpha_2\delta$  and  $\beta$ )( reviewed by De Waard, et al. (1997) in *Ion Channels*, Volume 4, edited by Narahashi, T. Plenum Press, New York). The  $\alpha_1$  subunit is the major pore-forming subunit and contains the voltage sensor and binding sites for calcium channel antagonists. The mainly extracellular  $\alpha_2$  is disulphide-linked to the transmembrane  $\delta$  subunit and both are derived from the same gene and are proteolytically cleaved *in vivo*. The  $\beta$  subunit is a non-glycosylated, hydrophilic protein with a high affinity of binding to a cytoplasmic region of the  $\alpha_1$  subunit. A fourth subunit,  $\gamma$ , is unique to L-type Ca channels expressed in skeletal muscle T-tubules. The isolation and characterization of  $\gamma$ -subunit-encoding cDNAs is described in US Patent No. 5,386,025 which is incorporated herein by reference.

Molecular cloning has revealed the cDNA and corresponding amino acid sequences of six different types of  $\alpha_1$  subunits ( $\alpha_{1A}$ ,  $\alpha_{1B}$ ,  $\alpha_{1C}$ ,  $\alpha_{1D}$ ,  $\alpha_{1E}$  and  $\alpha_{1S}$ ) and four types of  $\beta$  subunits ( $\beta_1$ ,  $\beta_2$ ,  $\beta_3$  and  $\beta_4$ )(reviewed in Stea, A., Soong, T.W. and Snutch, T.P. (1994) "Voltage-gated calcium channels." in *Handbook of Receptors and Channels*. Edited by R.A. North, CRC

Press.). PCT Patent Publication WO 95/04144, which is incorporated herein by reference, discloses the sequence and expression of  $\alpha_{1E}$  calcium channel subunits.

The different classes of  $\alpha 1$  and  $\beta$  subunits have been identified in different animals including, rat, rabbit and human and share a significant degree of amino acid conservation across species (for examples see: Castellano, et al. (1993) "Cloning and expression of a third calcium channel  $\beta$  subunit." *J. Biol. Chem.* 268: 3450-3455; Castellano, et al. (1993) "Cloning and expression of a neuronal calcium channel  $\beta$  subunit." *J. Biol. Chem.* 268: 12359-12366; Dubel, et al. (1992). "Molecular cloning of the  $\alpha_1$  subunit of an  $\omega$ -conotoxin-sensitive calcium channel." *Proc. Natl. Acad. Sci. (USA)* 89: 5058-5062; Fujita, et al.. (1993) "Primary structure and functional expression of the  $\omega$ -conotoxin-sensitive N-type calcium channel from rabbit brain." *Neuron* 10: 585-598; Mikami, et al.. (1989). "Primary structure and functional expression of the cardiac dihydropyridine-sensitive calcium channel." *Nature* 340: 230-233; Mori, et al. (1991) "Primary structure and functional expression from complementary DNA of a brain calcium channel." *Nature* 350: 398-402; Perez-Reyes, et al. (1992). "Cloning and expression of a cardiac/brain  $\beta$  subunit of the L-type calcium channel." *J. Biol. Chem.* 267: 1792-1797; Pragnell, et al. (1991). "Cloning and tissue-specific expression of the brain calcium channel  $\beta$ -subunit." *FEBS Lett.* 291: 253-258; Snutch, et al. (1991) "Distinct calcium channels are generated by alternative splicing and are differentially expressed in the mammalian CNS." *Neuron* 7: 45-57; Soong, et al. (1993) "Structure and functional expression of a member of the low voltage-activated calcium channel family." *Science* 260: 1133-1136; Tomlinson, et al. (1993) "Functional properties of a neuronal class C L-type channel." *Neuropharmacology* 32: 1117-1126; Williams, et al. (1992) "Structure and functional expression of  $\alpha 1$ ,  $\alpha 2$ , and  $\beta$  subunits of a novel human neuronal calcium channel subtype." *Neuron* 8: 71-84; Williams, et al. (1992) "Structure and functional expression of an  $\omega$ -conotoxin-sensitive human N-type calcium channel." *Science* 257: 389-395.

In some expression systems the  $\alpha_1$  subunits alone can form functional calcium channels although their electrophysiological and pharmacological properties can be differentially modulated by coexpression with any of the four  $\beta$  subunits. Until recently, the reported modulatory affects of  $\beta$  subunit coexpression were to mainly alter kinetic and voltage-dependent properties. More recently it has been shown that  $\beta$  subunits also play crucial roles in modulating channel activity by protein kinase A, protein kinase C and direct G-protein interaction. (Bourinet, et al. (1994) "Voltage-dependent facilitation of a neuronal  $\alpha_1C$  L-type calcium channel." *EMBO J.* 13: 5032-5039; Stea, et al. (1995) "Determinants of PKC-dependent modulation of a family of neuronal calcium channels." *Neuron* 15:929-940; Bourinet, et al. (1996) "Determinants of the G-protein-dependent opioid modulation of neuronal calcium channels." *Proc. Natl. Acad. Sci. (USA)* 93: 1486-1491.)

The electrophysiological and pharmacological properties of the calcium channels cloned to date can be summarized as shown in Table 1. While the cloned  $\alpha_1$  subunits identified to date correspond to several of the calcium channels found in cells, they do not account for all types of calcium conductances described in native cells. For example, they do not account for the various properties described for the heterogenous family described as T-type calcium channels. Furthermore, they do not account for novel calcium channels described in cerebellar granule cells or other types of cells. (Forti, et al (1993) "Functional diversity of L-type calcium channels in rat cerebellar neurons." *Neuron* 10: 437-450; Tottene, et al. (1996). "Functional diversity of P-type and R-type calcium channels in rat cerebellar neurons." *J. Neurosci.* 16: 6353-6363).

Because of the importance of calcium channels in cellular metabolism and human disease, it would be desirable to identify the remaining classes of  $\alpha_1$  subunits, and to develop expression systems for these subunits which would permit the study and characterization of these calcium channels, including the study of pharmacological modulators of calcium channel function. Thus, it is an object of the present invention to provide heretofor undisclosed calcium channels having novel  $\alpha_1$  subunits, including cell lines expressing these

TABLE 1

	$\omega$ -conotoxin GVIA	1,4- dihydropyridines	cadmium	$\omega$ -agatoxin IVA	$\omega$ -conotoxin MVIIIC	native $\text{Ca}^{2+}$ channel type
$\alpha_{1A}$	-	-	✓	✓	✓	P/Q-type
$\alpha_{1B}$	✓	-	✓	-	✓	N-type
$\alpha_{1C}$	-	✓	✓	-	-	L-type
$\alpha_{1D}$	-	✓	✓	-	-	L-type
$\alpha_{1E}$	-	-	✓	-	-	novel
$\alpha_{1S}$	-	✓	✓	-	-	L-type



new calcium channels. It is a further object of the present invention to provide a method for testing these novel calcium channels using such cell lines.

## SUMMARY OF THE INVENTION

The present invention provides sequences for a novel mammalian calcium channel subunits of T-type calcium channels, which we have labeled as  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  subunits. Knowledge of the sequences of these calcium channel subunits may be used in the development of probes for mapping the distribution and expression of the subunits in target tissues. In addition, these subunits, either alone or assembled with other proteins, can produce functional calcium channels, which can be evaluated in model cell lines to determine the properties of the channels containing the subunits of the invention. These cell lines can be used to evaluate the affects of pharmaceuticals and/or toxic substances on calcium channels incorporating  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  subunits

## BRIEF DESCRIPTION OF THE DRAWINGS

Figs. 1A and B show a comparison of the waveforms and current voltage relationship for  $\alpha_{1G}$ ;

Figs. 2A and B show a comparison of the waveforms and current voltage relationship for  $\alpha_{1I}$  calcium channels.

Fig. 3 shows a comparison of the steady state inactivation profiles of the  $\alpha_{1G}$  and  $\alpha_{1I}$  calcium channels.

Figs. 4A-C show a comparison of the inactivation kinetics of the  $\alpha_{1G}$  and  $\alpha_{1I}$  calcium channels.

## DETAILED DESCRIPTION OF THE INVENTION

The present invention includes the following aspects for which protection is sought:

5 (a) novel mammalian (including human) calcium channel subunits and DNA sequences encoding such subunits. Specifically, the invention encompasses an at least partially purified DNA molecule comprising a sequence of nucleotides that encodes an  $\alpha$  subunit of a T-type calcium channel, and such  $\alpha$  subunits *per se*. It will be appreciated that polymorphic variations may be made or may exist in the DNA of some individuals leading to minor deviations in the DNA or amino acids sequences from those shown which do not lead to any substantial alteration in the function of the calcium channel. Such variations, including variations which lead to substitutions of amino acids having similar properties are considered to be within the scope of the present invention. Thus, in one embodiment, the present application claims DNA molecules which encode  $\alpha_1$  subunits of mammalian T-type calcium channels, and which hybridize under conditions of medium (or higher) hybridization stringency with one or another of the specific sequences disclosed in this application. This level of hybridization stringency is generally sufficient given the length of the sequences involved to permit recovery of the subunits within the scope of the invention from mammalian DNA libraries.

10 (b) polynucleotide sequences useful as probes in screening human cDNA libraries for genes encoding these novel calcium channel subunits. These probes can also be used in histological assay to determine the tissue distribution of the novel calcium channel subunits.

15 (c) at least partially purified  $\alpha_1$  subunits and related peptides for mammalian T-type calcium channels. These proteins and peptides can be used to generate polyclonal or monoclonal antibodies to determine the cellular and subcellular distribution of T-type calcium channel subunits.

20 (d) eukaryotic cell lines expressing the novel calcium channel subunits. These cell lines can be used to evaluate compounds as pharmacological modifiers of the function of the novel calcium channel subunits.

(e) a method for evaluating compounds as pharmacological modifiers of the function of the novel calcium channel subunits using the cell lines expressing those subunits alone or in combination with other calcium channel subunits.

Further, since defects in the novel calcium channel subunits may be associated with a human genetic disease including, but not limited to; epilepsy, migraine, ataxia, schizophrenia, hypertension, arrhythmia, angina, depression, small lung carcinoma, Lambert-Eaton syndrome and Parkinson's disease; characterization of such associations and ultimately diagnosis of associated diseases can be carried out with probes which bind to the wild-type or defective forms of the novel calcium channels.

As used in the specification and claims of this application, the term "T-type calcium channel" refers to a voltage-gated calcium channel having a low activation voltage, generally less than -50 mV, and most commonly less than -60 mV. T-type calcium channels also exhibit comparatively negative steady-state inactivation properties and slow deactivation kinetics. The terms " $\alpha_1$  subunit" or " $\alpha_1$  calcium channel" refer to a protein subunit of a calcium channel which is responsible for pore formation and contains the voltage sensor and binding sites for calcium channel agonists and antagonists. Such subunits may be independently functional as calcium channels or may require the presence of other subunit types for complete functionality.

As used in the specification and claims of this application, the phrase "at least partially purified" refers to DNA or protein preparations in the which the specified molecule has been separated from adjacent cellular components and molecules with which it occurs in the natural state, either by virtue of performing a physical separation process or by virtue of making the DNA or protein molecule in a non-natural environment in the first place. The term encompasses cDNA molecules and expression vectors.

In accordance with the present invention, we have identified mammalian DNA sequences which code for novel T-type calcium channel  $\alpha_1$  subunits. These subunits are

believed to represent new types of  $\alpha_1$  subunits of mammalian voltage-dependent calcium channels which have been designated as types  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$ .

The novel  $\alpha_1$  subunits of the invention were identified by screening the *C. elegans* genomic DNA sequence data base for sequences homologous to previously identified mammalian calcium channel  $\alpha_1$  subunits. Specifically, the following twelve mammalian  $\alpha_1$  subunit sequences were used to screen the *C. elegans* genomic data bank:

rat brain $\alpha_{1A}$ :	GTCAAACTC AGGCCTTCTA CTGG	SEQ ID. No. 1
rat brain $\alpha_{1A}$ :	AACGTGTTCT TGGCTATCGC GGTG	SEQ ID. No. 2
rat brain $\alpha_{1B}$ :	GTGAAAGCAC AGAGCTTCTA CTGG	SEQ ID. No. 3
rat brain $\alpha_{1B}$ :	AACGTTTTCT TGGCCATTGC TGTG	SEQ ID. No. 4
rat brain $\alpha_{1C}$ :	GTAAATCCA ACGTCTTCTA CTGG	SEQ ID. No. 5
rat brain $\alpha_{1C}$ :	AATGTGTTCT TGGCCATTGC GGTG	SEQ ID. No. 6
rat brain $\alpha_{1D}$ :	GTGAAGTCTG TCACGTTTTA CTGG	SEQ ID. No. 7
rat brain $\alpha_{1D}$ :	AAGCTCTTCT TGGCCATTGC TGTA	SEQ ID. No. 8
rat brain $\alpha_{1E}$ :	GTCAAGTCGC AAGTGTTCTA CTGG	SEQ ID. No. 9
rat brain $\alpha_{1E}$ :	AATGTATTCT TGGCTATCGC TGTG	SEQ ID. No. 10
rat brain consensus #1 :	ATCTAYGCYR TSATYGGSAT G	SEQ ID. No. 11
rat brain consensus #2 :	ATGGACAAYT TYGASTAYTC	SEQ ID. No. 12

This search identified four distinct *C. elegans* cosmids that contain open reading frames (coding regions) that exhibit homology to mammalian calcium channel  $\alpha_1$  subunits:

cosmid and reading frame T02C5.5

cosmid and reading frame C48A7.1

cosmid and reading frame C54D2.5

cosmid and reading frame C27F2.3

Examination of the four *C. elegans* cosmid sequences by phylogeny analysis shows that two of these, T02C5.5 and C48A7.1, correspond closely with previously identified mammalian  $\alpha_1$  subunits. T02C5.5 appears to be an ancestral member related to the mammalian  $\alpha_{1A}$ ,  $\alpha_{1B}$  and  $\alpha_{1E}$  subunits. C48A7.1 appears to be an ancestral member related to the mammalian L-type channels encoded by  $\alpha_{1C}$ ,  $\alpha_{1D}$  and  $\alpha_{1S}$ . In contrast, the *C. elegans* cosmids C54D2.5 and C27F2.3 identify novel types of calcium channel  $\alpha_1$  subunits distinct from the other mammalian subtypes.

Mammalian counterparts of the *C. elegans* calcium channel  $\alpha_1$  subunit encoded by C54D2.5 were identified by screening of the GenBank expressed sequence tag (EST) data bank. This analysis identified a total of 13 mammalian sequences that exhibit some degree of DNA sequence and amino acid identity to C54D2.5, of which 8 are human sequences. (Table 2) Some of these sequences appear unlikely to encode novel calcium channel subunits because they either exhibit a significant degree of homology to previously identified mammalian  $\alpha_1$  subunits (for example, clones H06096 and H14053) or exhibit homology in a region not considered to be diagnostic of calcium channel  $\alpha_1$  subunits specifically as opposed to other types of ion channel molecules in general (for example, clone D20469). One of the five remaining sequences was evaluated and appears to encode a sodium channel. Four sequences (H55225, H55617, H55223, and H55544), however, encode what are believed to be previously unidentified calcium channel  $\alpha_1$  subunits. For these subunits, the amino acid sequences closely match that of known calcium channel subunits in conserved regions but are sufficiently different to indicate that they do not encode previously identified mammalian calcium channel  $\alpha_1$  subunits,  $\alpha_{1A}$ ,  $\alpha_{1B}$ ,  $\alpha_{1C}$ ,  $\alpha_{1D}$ ,  $\alpha_{1E}$ , or  $\alpha_{1S}$ . The expected amino acid sequence closely matches but is not identical to amino acid sequences in these known calcium channel subunits.

Table 2

Query = C54D2.5 CE02562 CALCIUM CHANNEL ALPHA-1 SUBUNIT LG:6

Database: Non-redundant Database of GenBank EST Division

824,500 sequences; 302,742,428 total letter

Sequences producing High-scoring Segment Pairs: Frame Score P(N)

gb AA183990 AA183990	ms53e02.r1 Life Tech mouse embry...	+1	108	1.8e-24
gb H55225 H55225	CHR220164 Homo sapiens genomic c...	+1	136	2.5e-10
dbj D68412 CELK131B1F	C.elegans cDNA clone yk131b1 : 5...	+3	117	1.7e-06
gb R75128 R75128	MDB1075 Mouse brain, Stratagene ...	+3	113	7.2e-06
gb H55617 H55617	CHR220556 Homo sapiens genomic c...	+2	102	2.8e-05
emb F07776 HSC2HD061	H. sapiens partial cDNA sequence...	+3	100	0.00057
gb W76774 W76774	me84e08.r1 Soares mouse embryo N...	+2	98	0.0012
gb H06096 H06096	y177e01.r1 Homo sapiens cDNA clo...	+3	98	0.0015
gb H14053 H14053	ym65d10.r1 Homo sapiens cDNA clo...	+2	91	0.0036
gb H55223 H55223	CHR220162 Homo sapiens genomic c...	+2	87	0.0039
dbj D35703 CELK024D9F	C.elegans cDNA clone yk24d9 : 5'...	+3	74	0.046
dbj D20469 HUMGS01443	Human HL60 3'directed MboI cDNA, ...	-2	66	0.91
gb H55544 H55544	CHR220483 Homo sapiens genomic c...	+1	65	0.98

The four sequences (H55225, H55617, H55223, and H55544) are found on human chromosome 22, and are now believed to all be part of the same gene encoding the novel human calcium channel subunit  $\alpha_{11}$ .

The sequences of the four selected sequences and the references from which they are taken are given as follows:

H55225 SOURCE human clone=C22\_207 primer=T3 library=Chromosome 22  
exon

Trofatter, et al., *Genome Res.* 5 (3): 214-224 (1995)

SEQ ID No. 13

1 GTGATCACTC TGAAGGCTG GGTGGAGATC ATGTACTACG TGATGGATGC TCACTCCTTC  
61 TACAACTTCA TCTACTTCAT CCTGCTTATC ATACCCCTCT TGCCTTGCAC CCCATATGGT

121 CTTCCCAGAG TGAGCTCATC CACCTCGTCA TGCCTGACTC GACGTTCA

H55617 SOURCE human clone=C22\_757 primer=T3 library=Chromosome 22  
exon

Trofatter, et al., *Genome Res.* 5 (3): 214-224 (1995)

SEQ ID No. 14

1 GATGGTCGAG TACTCCCTGG ACCTTCAGAA CATCAACCTG TCAGCCATCC GCACCGTGCG  
61 CGTCCTGAGG CCCCTCAAAG CCATCAACCG CGTGCCCA

H55223 SOURCE human clone=C22\_204 primer=T3 library=Chromosome 22  
exon

Trofatter, et al, *Genome Res.* 5 (3): 214-224 (1995)

SEQ ID No. 15

1 CATGCTGGTG ATCCTGCTGA ACTGCGTGAC ACTTGGCATG TACCAGCCGT GCGACGACAT  
61 GGACTGCCTG TCCGACCGCT GCAAGATCCT GCAG

H55544 SOURCE human clone=C22\_651 primer=T3 library=Chromosome 22  
exon

Trofatter, et al, *Genome Res.* 5 (3): 214-224 (1995)

SEQ ID No. 16

1 GTATCTCTGG TTACTTTAGT AGCCAACACT CTTGGCTACT CAGACCTTGG TCCCATTA  
61 TCCCTGCGAA CCTTGAGAGC ACTAAGACCT CTAAGAGCTT TGTCTAGATT TGAAGGAATG  
121 AGG

A search of the Sanger Genome Sequencing Center (Cambridge, U.K.) and the Washington University Genome Sequencing Center (St. Louis, MO) sequences in progress revealed a Bacterial Artificial Chromosome (BAC) sequence (bK206c7) that contained matches to the *C. elegans* cosmid open reading frame, C54D2.5, and to the four human

chromosome 22 ESTs, H55225, H55617, H55223, H55544. The *C. elegans* C54D2.5 cosmid sequence and the human EST sequences were then used to compare the translation of the bK206c7 BAC genomic sequence in all 6 reading frames. The analysis was performed using the graphical program Dotter (Eric Sohnhammer, NCBI). The analysis revealed a series of potential coding regions on one strand of the bK206c7 BAC sequence. These were subsequently translated in all 3 reading frames and the potential splice junctions identified. The translated sequence of this longer DNA fragment which is part of the human  $\alpha_{11}$  subunit gene is given by Seq. ID Nos. 17 and 18.

Using the sequence information from the four EST's, a full length gene can be recovered using any of several techniques. Polynucleotide probes having a sequence which corresponds to or hybridizes with the EST sequences or a distinctive portion thereof (for example oligonucleotide probes having a length of 18 to 100 nucleotides) can be used to probe a human cDNA library for identification of the full length DNA encoding the  $\alpha_{11}$  subunits. The process of identifying cDNAs of interest using defined probes is well known in the art and is, for example, described in International Patent Publication No. WO95/04144, which is incorporated herein by reference. This process generally involves screening bacterial hosts (e.g. *E. coli*) harboring the library plasmids or infected with recombinant lambda phage with labeled probes, e.g. radiolabeled with  $^{32}\text{P}$ , and selection of colonies or phage which bind the labeled probe. Each selected colony or phage is grown up, and the plasmids are recovered. Human cDNAs are recovered from the plasmids by restriction digestion, or can be amplified, for example by PCR. The recovered cDNA can be sequenced, and the position of the calcium channel subunit-encoding region further refined, although neither process is not necessary to the further use of the cDNA to produce cell lines expressing the novel calcium channel subunits.

Longer portions of DNA-encoding the novel calcium channel subunits of the invention can also be recovered by PCR cloning techniques using primers corresponding to or based upon the EST sequences. Using this technique to identify relevant sequences within a human



brain total RNA preparation confirmed that the novel  $\alpha_{11}$  calcium channel subunit is present in human brain. Subcloning of the 567 nt PCR product (Seq. ID No. 19, amino acids Seq. ID No. 20) and subsequent sequencing thereof showed that this product corresponds to the derived sequence from the bK206c7 BAC genomic sequence, the nucleotide sequence of which is given as SEQ ID No. 17 (amino acid sequence Seq. ID No. 18). The same experiment was performed using a rat brain RNA preparation and resulted in recovery of a substantially identical PCR product. (SEQ ID. No. 21). The protein encoded by the rat PCR product (SEQ ID No. 22) is 96% identical to the human PCR product (Seq. ID No. 20).

These sequences, which encode a partial subunit can be used as a basis for constructing full length human or rat  $\alpha_{11}$  clones. Briefly, the subcloned  $\alpha_{11}$  PCR product is radiolabeled by random hexamer priming according to standard methods (See, Sambrook, J., Fritsch, E.F. and Maniatis, T. (1989) *Molecular Cloning, A Laboratory Manual*. Cold Spring Harbor Press) and used to screen commercial human brain cDNA libraries (Stratagene, La Jolla, CA). The screening of cDNA libraries follows standard methods and includes such protocols as infecting bacteria with recombinant lambda phage, immobilizing lambda DNA to nitrocellulose filters and screening under medium hybridization stringency conditions with radiolabeled probe. cDNA clones homologous to the probe are identified by autoradiography. Positive clones are purified by sequential rounds of screening.

Following this protocol, most purified cDNA's are likely to be partial sequence clones due the nature of the cDNA library synthesis. Full length clones are constructed from cDNA's which overlap in DNA sequence. Restriction enzyme sites which overlap between cDNAs are used to ligate the individual cDNA's to generate a full-length cDNA. For subsequent heterologous expression, the full-length cDNA is subcloned directly into an appropriate vertebrate expression vector, such as pcDNA-3 (Invitrogen, San Diego, CA) in which expression of the cDNA is under the control of a promoter such as the CMV major intermediate early promoter/enhancer. Other suitable expression vectors include, for example, pMT2, pRC/CMV, pcDNA3.1 and pCEP4.

Following these protocols, as described more fully in Example 4, full length mammalian  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  calcium channel subunit cDNAs were isolated by using the 567 base pair human fragment (Seq. ID No. 19) to screen a rat brain cDNA library. Sequencing of the recovered sequences identified the three distinct classes of calcium channel subunits which have been demoninated herein as  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  subunits. For each class of subunit, complete sequencing of the largest cDNA confirmed that it represented only a portion of the predicted calcium channel coding region. Complete sequences for the three new subunits were obtained by rescreening the rat brain cDNA library with probes derived from the partial length cDNAs to obtain overlapping segments. These segments were combined to form a complete gene by restriction digestion and ligation. The complete cDNA sequences of the rat  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  subunits are given by Sequence ID Nos. 23, 25 and 27, respectively. Corresponding amino acid sequences are given by Sequence ID Nos. 24, 26 and 28. The same techniques are employed to recover human sequences by screening of a human or other mammalian library. Thus, for example, partial length human sequences for  $\alpha_{1G}$  and  $\alpha_{1H}$  T-type calcium channels have been recovered using the same probe (Seq. ID No. 19) and the full length rat  $\alpha_{1I}$  cDNA (Seq. ID. No. 27) has been used to recover a partial length DNA encoding a human  $\alpha_{1I}$  T-type calcium channel. The DNA and amino acid sequences for these partial length human calcium channels are given by Seq. ID Nos. 30-35.

Once the full length cDNA is cloned into an expression vector, the vector is then transfected into a host cell for expression. Suitable host cells include *Xenopus* oocytes or mammalian cells such as human embryonic kidney cells as described in International Patent Publication No. WO 96/39512 which is incorporated herein by reference and Ltk cells as described in US Patent No. 5,386,025 which is incorporated herein by reference. Transfection into host cells may be accomplished by microinjection, lipofection, glycerol shock, electroporation calcium phosphate or particle-mediated gene transfer. The vector may also be transfected into host cells to provide coexpression of the novel  $\alpha_1$  subunits with a  $\beta$  and/or an  $\alpha_2\delta$  subunit.

To confirm that the three full length cDNAs (sequence ID Nos. 23, 25 and 27) encoded function calcium channels, the  $\alpha_{1G}$  and  $\alpha_{1H}$  cDNAs were transiently transfected into human embryonic kidney cells and evaluated using electrophysiological recording techniques. As described in more detail in Example 5 below, and as illustrated in Figs. 1-4), the results are consistent with a role of these subunits in native T-type channels in nerve, muscle and endocrine cells.

The resulting cell lines expressing functional calcium channels including the novel  $\alpha_1$  subunits of the invention can be used test compounds for pharmacological activity with respect to these calcium channels. Thus, the cell lines are useful for screening compounds for pharmaceutical utility. Such screening can be carried out using several available methods for evaluation of the interaction, if any, between the test compound and the calcium channel. One such method involves the binding of radiolabeled agents that interact with the calcium channel and subsequent analysis of equilibrium binding measurements including but not limited to, on rates, off rates,  $K_d$  values and competitive binding by other molecules. Another such method involves the screening for the effects of compounds by electrophysiological assay whereby individual cells are impaled with a microelectrode and currents through the calcium channel are recorded before and after application of the compound of interest. Another method, high-throughput spectrophotometric assay, utilizes the loading the cell lines with a fluorescent dye sensitive to intracellular calcium concentration and subsequent examination of the effects of compounds on the ability of depolarization by potassium chloride or other means to alter intracellular calcium levels. Compounds to be tested as agonists or antagonists of the novel  $\alpha_{1H}$  calcium channel subunits are combined with cells that are stably or transiently transformed with a DNA sequence encoding the  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  calcium channel subunits of the invention and monitored using one of these techniques.

DNA fragments with sequences given by SEQ ID Nos. 13-17 and 19, or polynucleotides with the complete coding sequences as given by Sequence ID Nos. 23, 25 and 27 or distinctive portions thereof which do not exhibit non-discriminatory levels of homology

with other types of calcium channel subunits may also be used for mapping the distribution of  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  calcium channel subunits within a tissue sample. This method follows normal histological procedures using a nucleic acid probe, and generally involves the steps of exposing the tissue to a reagent comprising a directly or indirectly detectable label coupled to a selected DNA fragment, and detecting reagent that has bound to the tissue. Suitable labels include fluorescent labels, enzyme labels, chromophores and radio-labels.

### EXAMPLE 1

In order to isolate novel human calcium channel  $\alpha_1$  subunits using standard molecular cloning protocols, synthetic DNA probes are prepared, radiolabeled with  $^{32}\text{P}$  and utilized to screen human cDNA libraries commercially available in lambda phage vectors (Stratagene, La Jolla, CA) based on the human DNA sequences for H55225, H55617, H55223, and H55544. DNA fragments with the sequence of sequence ID Nos 17 and 19 may also be used for this purpose. Positive phage are purified through several rounds of screening involving immobilizing the phage DNA on nitrocellulose filters, hybridizing with the radiolabeled probe, washing off of excess probe and then selection of clones by autoradiography. Clones identified by this approach are expected to be partial length clones due to the nature of cDNA library synthesis and several rounds of screening for each calcium channel type may be necessary to obtain full-length clones.

To characterize the clones, double stranded plasmid DNA is prepared from the identified clones and the sequences are determined using  $^{35}\text{S}$  dATP, Sequenase and standard gel electrophoresis methods. Regions of similarity and regions of overlap are determined by comparison of each cDNA sequence.

Full-length clones are constructed by ligating overlapping cDNA fragments together at common restriction enzyme sites. The full-length clones are subsequently inserted into vectors suitable for expression in vertebrate cells (e.g. pMT2, pRC/CMV, pcDNA3.1, pCEP4,

pREP7) by ligation into restriction sites in the vector polylinker region which is downstream of the promoter used to direct cDNA expression.

DNA encoding the novel calcium channels can be stably or transiently introduced into eukaryotic cells (e.g. human embryonic kidney, mouse L cells, chinese hamster ovary, etc) by any number of available standard methods. Stable transfection is achieved by growing the cells under conditions that promote growth of cells expressing a marker gene which is contained in the expression vector (e.g. dihydrofolate reductase, thymidine kinase, or the like). The heterologous DNA encoding the human calcium channel may be integrated into the genome or may be maintained as an episomal element.

Expression of the human calcium channel in transfected cells may monitored by any number of techniques, including Northern blot for RNA analysis, Southern blot for cDNA detection, electrophysiological assay for calcium channel function, the binding of radiolabeled agents thought to interact with the calcium channel, and fluorescent assay of dyes sensitive to intracellular calcium concentration.

## EXAMPLE 2

### Heterologous Expression of Mammalian $\alpha_{11}$ Calcium Channels in Cells

#### A. Transient Transfection in Mammalian Cells

Host cells, such as human embryonic kidney cells, HEK 293 (ATCC# CRL 1573) are grown in standard DMEM medium supplemented with 2 mM glutamine and 10% fetal bovine serum. HEK 293 cells are transfected by a standard calcium-phosphate-DNA co-precipitation method using a full-length mammalian  $\alpha_{11}$  calcium channel cDNA (for example, Seq. ID. No. 27) in a vertebrate expression vector (for example see Current protocols in Molecular Biology). The  $\alpha_{11}$  calcium channel cDNA may be transfected alone or in combination with other cloned subunits for mammalian calcium channels, such as  $\alpha_{2\delta}$  and  $\beta$  subunits, and also with clones for marker proteins such the jellyfish green fluorescent protein.

Electrophysiological Recording: After an incubation period of from 24 to 72 hrs the culture medium is removed and replaced with external recording solution (see below). Whole cell patch clamp experiments are performed using an Axopatch 200B amplifier (Axon Instruments, Burlingame, CA) linked to an IBM compatible personal computer equipped with pCLAMP software. Microelectrodes are filled with 3 M CsCl and have typical resistances from 0.5 to 2.5 MΩ. The external recording solution is 20 mM BaCl<sub>2</sub>, 1 mM MgCl<sub>2</sub>, 10 mM HEPES, 40 mM TEACl, 10 mM Glucose, 65 mM CsCl, (pH 7.2). The internal pipette solution is 105 mM CsCl, 25 mM TEACl, 1 mM CaCl<sub>2</sub>, 11 mM EGTA, 10 mM HEPES (pH 7.2). Currents are typically elicited from a holding potential of -100 mV to various test potentials. Data are filtered at 1 kHz and recorded directly on the harddrive of a personal computer. Leak subtraction is carried out on-line using a standard P/5 protocol. Currents are analyzed using pCLAMP versions 5.5 and 6.0. Macroscopic current-voltage relations are fitted with the equation  $I = \{1/(1+\exp(-(V_m-V_h)/S))\} \times G - (V_m-E_{rev})$ , where  $V_m$  is the test potential,  $V_h$  is the voltage at which half of the channels are activated, and S reflects the steepness of the activation curve and is an indication of the effective gating charge movement. Inactivation curves are normalized to 1 and fitted with  $I = (1/1 + \exp((V_m-V_h)/S))$  with  $V_m$  being the holding potential. Single channel recordings are performed in the cell-attached mode with the following pipette solution (in mM): 100 BaCl<sub>2</sub>, 10 HEPES, pH 7.4 and bath solution: 100 KCl, 10 EGTA, 2 MgCl<sub>2</sub>, 10 HEPES, pH 7.4.

#### **B. Transient Transfection in Xenopus Oocytes**

Stage V and VI Xenopus oocytes are prepared as described by Dascal et al (1986), Expression and modulation of voltage-gated calcium channels after RNA injection into Xenopus oocytes. Science 231:1147-1150. After enzymatic dissociation with collagenase, oocytes nuclei are microinjected with the human  $\alpha_{1I}$  calcium channel cDNA expression vector construct (approximately 10 ng DNA per nucleus) using a Drummond nanoject apparatus. The  $\alpha_{1I}$  calcium channel may be injected alone, or in combination with other mammalian

calcium channel subunit cDNAs, such as the  $\alpha 2\text{-}\delta$  and  $\beta 1b$  subunits. After incubation from 48 to 96 hrs macroscopic currents are recorded using a standard two microelectrode voltage-clamp (Axoclamp 2A, Axon Instruments, Burlingame, CA) in a bathing medium containing (in mM): 40 Ba(OH)<sub>2</sub>, 25 TEA-OH, 25 NaOH, 2 CsOH, 5 HEPES (pH titrated to 7.3 with methan-sulfonic acid). Pipettes of typical resistance ranging from 0.5 to 1.5 m $\Omega$  are filled with 2.8M CsCl, 0.2M CsOH, 10mM HEPES, 10mM BAPTA free acid. Endogenous Ca (and Ba) -activated Cl currents are suppressed by systematically injecting 10-30 nl of a solution containing 100mM BAPTA-free acid, 10mM HEPES (pH titrated to 7.2 with CsOH) using a third pipette connected to a pneumatic injector. Leak currents and capacitive transients are subtracted using a standard P/5 procedure.

### EXAMPLE 3

#### Construction of Stable Cell Lines Expressing Mammalian $\alpha_{11}$ Calcium Channels

Mammalian cell lines stably expressing human  $\alpha_{11}$  calcium channels are constructed by transfecting the  $\alpha_{11}$  calcium channel cDNA into mammalian cells such as HEK 293 and selecting for antibiotic resistance encoded for by an expression vector. Briefly, a full-length mammalian  $\alpha_{11}$  calcium channel cDNA (for example Seq. ID No. 27) subcloned into a vertebrate expression vector with a selectable marker, such as the pcDNA3 (InvitroGen, San Diego, CA), is transfected into HEK 293 cells by calcium phosphate coprecipitation or lipofection or electroporation or other method according to well known procedures (Methods in Enzymology, Volume 185, Gene Expression Technology (1990) Edited by Goeddel, D.V.). The  $\alpha_{11}$  calcium channel may be transfected alone, or in combination with other mammalian calcium channel subunit cDNAs, such as the  $\alpha 2\text{-}\delta$  and  $\beta 1b$  subunits, either in a similar expression vector or other type of vector using different selectable markers. After incubation for 2 days in nonselective conditions, the medium is supplemented with Geneticin (G418) at a concentration of between 600 to 800 ug/ml. After 3 to 4 weeks in this medium, cells which are resistant to G418 are visible and can be cloned as isolated colonies using standard cloning

rings. After growing up each isolated colony to confluency to establish cell lines, the expression of  $\alpha_{11}$  calcium channels can be determined at with standard gene expression methods such as Northern blotting, RNase protection and reverse-transcriptase PCR.

The functional detection of  $\alpha_{11}$  calcium channels in stably transfected cells can be examined electrophysiologically, such as by whole patch clamp or single channel analysis (see above). Other means of detecting functional calcium channels include the use of radiolabeled  $^{45}\text{Ca}$  uptake, fluorescence spectroscopy using calcium sensitive dyes such as FURA-2, and the binding or displacement of radiolabeled ligands that interact with the calcium channel.

#### EXAMPLE 4

In order to recover full-length mammalian sequences for novel calcium channels, the 567 base pair partial length human brain  $\alpha_{11}$  cDNA was gel-purified, radio-labelled with  $^{32}\text{P}$  dATP and dCTP by random priming (Feinberg et al., 1983, *Anal. Biochem.* 132: 6-13) and used to screen a rat brain cDNA library constructed in the phase vector Lambda Zapp II. (Snutch et al., 1990, *Proc Natl Acad Sci (USA)* 87: 3391-3395). Screening was carried out at 62°C in 5XSSPE (1XSSPE= 0.18 M NaCl; 1mM EDTA; 10 mM sodium phosphate, pH=7.4 0.3% SDS, 0.2 mg/ml denatured salmon sperm DNA). Filters were washed at 62°C in 0.2X SSPE/0.1% SDS. After three rounds of screening and plaque purification, positive phages were transformed into Bluescript phagemids (Stratagene, La Jolla, CA) by *in vivo* excision.

Double stranded DNA sequencing on the recombinant phagemids was performed using a modified dideoxynucleotide protocol (Biggin et al., 1983, *Proc Natl Acad Sci (USA)* 80:3963-3965) and Sequenase version 2.1 (United States Biochemical Corp.). DNA sequencing identified three distinct classes of calcium channel  $\alpha_1$  subunits: designated as  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{11}$  calcium channel subunits.

For each class of calcium channel  $\alpha_1$  subunit, the largest cDNA was completely sequenced and determined to represent only a portion of the predicted calcium channel



coding region. In order to isolate the remaining portions of  $\alpha_{1G}$  and  $\alpha_{1I}$  calcium channel subunits, the  $\alpha_{1G}$  clone was digested with HindIII and SpeI. The resulting 540 base pair fragment was gel purified, radiolabeled with  $^{32}P$  dATP and dCTP by random priming and used to rescreen the rat brain cDNA library as described above. The sequence of the 540 base pair  $\alpha_{1G}$  screening probe used is given by Seq. ID No. 29. Other sequences spanning regions of distinctiveness within the sequences for the subunits could also be employed.

Double-stranded DNA sequencing of the purified recombinant phagemids showed that additional  $\alpha_{1G}$ ,  $\alpha_{1H}$  and  $\alpha_{1I}$  calcium channel subunit cDNAs overlapped with the original partial length cDNAs and together encoded complete protein coding regions as well as portions of their respective 5' and 3' non-coding untranslated regions.

To recover further human sequences for the novel  $\alpha_{1G}$  and  $\alpha_{1H}$  calcium channels, the 567 base pair partial length human brain  $\alpha_{1I}$  cDNA (Seq. 19) was radio-labelled with  $^{32}P$  dATP and dCTP by random priming and used to screen a commercial human thalamus cDNA library (Clontech). Hybridization was performed overnight at 65 °C in 6 X SSPE; 0.3% SDS; 5X Denhardt's. Filters were washed at 65 °C in 0.1 X SSPE/ 0.3% SDS. After four rounds of screening and plaque purification, positive phages were selected, DNA prepared and the insert cDNA excised from the lambda vector by digestion with Eco R1 restriction enzyme. The excised cDNA was subcloned into the plasmid Bluescript KS (Stratagene, La Jolla, CA) and the DNA sequence determined using a modified dideoxynucleotide protocol and Sequenase version 2.1. The partial length  $\alpha_{1G}$  cDNA isolated consisted of 2212 base pairs of which 279 base pairs were 5' noncoding and 1,933 base pairs were coding region representing 644 amino acids (Seq. ID Nos. 30, 31). The partial  $\alpha_{1H}$  cDNA isolated consisted of 1,608 base pairs of which 53 base pairs were 5' noncoding and 1,555 were coding region representing 518 amino acids (Seq. ID Nos. 32, 33).

To recover further human sequences for the novel  $\alpha_{1I}$  calcium channel, the full-length rat brain  $\alpha_{1I}$  cDNA (Seq. 27) was radio-labelled  $^{32}P$  dATP and dCTP by random priming and used to screen a commercial human hippocampus cDNA library (Stratagene). Hybridization

was performed overnight at 65 °C in 6 X SSPE; 0.3% SDS; 5X Denhardt's. Filters were washed at 65 °C in 0.1 X SSPE/ 0.3% SDS. After four rounds of screening and plaque purification, positive phages were transformed into Bluescript phagemids (Stratagene, LA Jolla, CA) by *in vitro* excision. The excised cDNA DNA sequence was determined using a modified dideoxynucleotide protocol and Sequenase version 2.1. The partial  $\alpha_{1I}$  cDNA isolated consisted of 1,080 base pairs of coding region representing 360 amino acids (Seq. ID Nos. 34, 35).

#### EXAMPLE 5

Double-stranded DNA sequencing of the purified recombinant phagemids showed that additional  $\alpha_{1G}$  and  $\alpha_{1I}$  calcium channel cDNAs overlapped with the original partial length cDNAs and together encoded complete protein coding regions as well as portions of their respective 5' and 3' non-coding untranslated regions. (Seq. ID Nos. 23 and 27, respectively) In addition to the  $\alpha_{1G}$  and  $\alpha_{1I}$  calcium channel classes, DNA sequencing of the recombinant phagemids also identified a third class of calcium channel  $\alpha_1$  subunit: designated as the  $\alpha_{1H}$  calcium channel subunit. The partial length  $\alpha_{1H}$  calcium channel cDNAs overlapped and together encoded a complete  $\alpha_{1H}$  coding region as well as portions of the 5' and 3' untranslated regions (Seq. ID. No. 25).

Electrophysiological studies were performed on transiently-transfected human embryonic kidney cells (HEK-tsa201) prepared using the general protocol of Example 2. Transfection was carried out by standard calcium phosphate precipitation. (Okayama et al., 1991, *Methods in Molec. Biol.*, Vol. 7, ed. Murray, E.J.). For maintenance, HEK-tsa201 cells were cultured until approximately 70% confluent, the media removed and cells dispersed with trypsin and gentle trituration. Cells were then diluted 1:10 in culture medium (10% FBS, DMEM plus L-glutamine, pen-stp) warmed to 37°C and plated onto tissue culture dishes. For transient transfection, 0.5 mM  $\text{CaCl}_2$  was mixed with a total of 20  $\mu\text{g}$  of DNA (consisting of 3 $\mu\text{g}$  of either rat brain  $\alpha_{1G}$  or  $\alpha_{1I}$  calcium channel cDNA, 2  $\mu\text{g}$  of CD8 plasmid marker, and

15  $\mu$ g of Bluescript plasmid carrier DNA). The DNA mixture was mixed thoroughly and then slowly added dropwise to 0.5 ml of 2 times HeBS (274 mM NaCl, 20mM D-glucose, 10mM KCl, 1.4 mM  $\text{Na}_2\text{HPO}_4$ , 40 mM Hepes (pH=7.05). After incubation at room temperature for 20 min, 100  $\mu$ l of the DNA mixture was slowly added to each dish of HEK-tsa201 cells and then incubated at 37°C for 24 to 48 hours in a tissue culture incubator (5%  $\text{CO}_2$ ).

Positive transfectant cells were identified visually by addition of 1  $\mu$ l of mouse CD8 (Lyt2) Dynabeads directly to the recording solution and gentle swirling to mix. Whole cell patch clamp readings were carried out with an Axopatch 200A amplifier (Axon Instruments) as described previously. (Zamponi et al., 1997, *Nature* 385: 442-446). The external recording solution was 2 mM  $\text{CaCl}_2$ , 1 mM  $\text{MgCl}_2$ , 10 mM HEPES, 40 mM TEA-Cl, 10 mM glucose, 92 mM CsCl, pH=7.2 with TEA-hydroxide. The internal pipette solutions was 105 mM CsCl, 25 mM TEA-Cl, 1mM  $\text{CaCl}_2$ , 11 mM EGTA, 10 mM HEPES, pH 7.2 with NaOH.

For determination of current-voltage (I-V) relationships, cells were held at a resting potential of -100 mV and then stepped to various depolarizing test potentials. For steady-state inactivation, cells were held at various potentials for 20 sec. and currents recorded during a subsequent test pulse to the peak potential of the I-V. Leak currents and capacitive transients were subtracted using a P/5 procedure.

Figs. 1-4 show the results obtained for HEK cells transfected with and expressing the cDNA of sequences ID Nos. 23 and 27, which correspond to the subunits designated as  $\alpha_{1G}$  and  $\alpha_{1I}$ . Figs. 1A and B and 2A and B shows a comparison of the waveforms and current-voltage relationship for the two channel subunit types. In the presence of recording solution containing 2mM  $\text{Ca}^{2+}$ , both the  $\alpha_{1G}$  and  $\alpha_{1I}$  channel subunits exhibit activation properties consistent with native T-type calcium currents. Figs 1 A and 2A show the subunit calcium current from a cell held at -120 mV and depolarized to a series of test potentials. Figs 1B and 2B show the magnitude of the calcium current. From a holding potential of -110 mV, both channel first activate at approximately -70 mV and peak currents are obtained between -40 and -50 mV. Upon depolarization to various test potentials, the current waveforms of the  $\alpha_{1G}$

and  $\alpha_{11}$  calcium channels exhibit an overlapping pattern characteristic of native T-type channels in nerve, muscle and endocrine cells.

Fig. 3 shows steady-state inactivation profiles for the  $\alpha_{1G}$  and  $\alpha_{11}$  calcium channels in HEK 293 cells transiently transformed with full length cDNAs (SEQ ID Nos 23 or 27) for  $\alpha_{1G}$  or  $\alpha_{11}$  subunits. Steady state inactivation properties were determined by stepping from -120 mV to prepulse holding potentials between -120 mV and -50 mV for 15 sec., prior to a test potential of -30 mV. The data are plotted as normalized whole cell current versus prepulse holding potential and show that  $\alpha_{1G}$  exhibits a  $V_{50}$  of approximately -85 mV and  $\alpha_{11}$  a  $V_{50}$  of approximately -93 mV. Thus, consistent with native T-type calcium channels, both of the  $\alpha_{1G}$  and  $\alpha_{11}$  calcium channels exhibit pronounced steady-state inactivation at negative potentials.

Figs. 4A-C show a comparison of the voltage-dependent deactivation profiles of the  $\alpha_{1G}$  and  $\alpha_{11}$  calcium channels. HEK 293 cells were transiently transfected with either an  $\alpha_{1G}$  or  $\alpha_{11}$  subunit cDNA (Seq. ID No. 23 or 27). The deactivation properties of  $\alpha_{1G}$  were determined by stepping from a holding potential of -100 mV to -40mV for 9 msec, and then to potentials between -120 mV and -45 mV. The deactivation properties of  $\alpha_{11}$  were determined by stepping from a holding potential of -100 mV to -40 mV for 20 msec, and then to potentials between -120 mV and -45 mV. Both channels exhibit slow deactivation kinetics compared to typical high-threshold channels, and is consistent with the  $\alpha_{1G}$  and  $\alpha_{11}$  subunits being subunits for T-type calcium channels

What is claimed is:

1           1.     An at least partially purified DNA molecule comprising a sequence of  
2 nucleotides that encodes an  $\alpha_1$  subunit of a mammalian T-type calcium channel.

1           2.     The DNA molecule of claim 1, wherein the sequence of nucleotides is selected  
2 from sequences of nucleotides encoding a protein including the sequence of amino acids set  
3 forth in SEQ ID. No. 18, 20, 24, 26, 28, 31, 33, or 35 and sequences of nucleotides that  
4 hybridize under conditions of medium hybridization stringency to DNA encoding a protein  
5 including the sequence set forth in SEQ ID No. 18, 20, 24, 26, 28, 31, 33, 35.

1           3.     The DNA molecule of Claim 1, wherein the calcium channel is a human  
2 calcium channel.

1           4.     The DNA molecule of claim 1, further comprising a promoter region effective  
2 to promote expression of the  $\alpha_1$  subunit of a mammalian T-type calcium channel when the  
3 DNA molecule is transfected into a vertebrate cell.

1           5.     The DNA molecule of claim 1, having the sequence as set forth in Seq. ID. No.  
2 23, 25 or 27.

1           6.     The DNA molecule of claim 1, wherein the molecule comprises a region  
2 consisting of the sequence as set forth in Seq. ID. No. 30, 32 or 34.

1           7.     An at least partially purified  $\alpha_1$  subunit of a mammalian T-type calcium  
2 channel.

1           8.     The  $\alpha_1$  subunit of claim 7 , wherein the subunit has the sequence as set forth in  
2     Seq. ID No. 24, 26 or 28 .

1           9.     The  $\alpha_1$  subunit of claim 7 , wherein the subunit comprises a region consisting  
2     of the sequence as set forth in Seq. ID. No. 31, 33 or 35.

1           10.    A eukaryotic cell transiently or stably transformed with the vertebrate  
2     expression vector comprising a sequence of nucleotides that encodes an  $\alpha_1$  subunit of a  
3     mammalian T-type calcium channel, wherein the cell expresses the  $\alpha_1$  subunit of a  
4     mammalian T-type calcium channel.

1           11.    The cell of claim 10 , wherein the sequence of nucleotides is selected from  
2     sequences of nucleotides encoding a protein including the sequence of amino acids set forth in  
3     SEQ ID. No. 18, 20, 24, 26, 28, 31, 33, or 35, and sequences of nucleotides that hybridize  
4     under conditions of medium hybridization stringency to DNA encoding a protein including  
5     the sequence set forth in SEQ ID No. 18, 20, 24, 26, 28, 31, 33, or 35.

1           12.    The cell of claim 10 , wherein the calcium channel is a human calcium  
2     channel.

1           13.    The cell of claim 10 , wherein the sequence of nucleotides has the sequence as  
2     set forth in Seq. ID. No. 23, 25 or 27 .

1           14.    The cell of claim 10 , wherein the sequence of nucleotides comprises a region  
2     consisting of the sequence as set forth in Seq. ID. No. 30, 32 or 34.

1           15.    The cell of claim 10 , wherein the sequence of nucleotides has the sequence as  
2 set forth in Seq. ID. No. 27.

1           16.    The cell of claim 10 , wherein the cell is further transformed with and  
2 expresses an  $\alpha_2\delta$  or a  $\beta$  calcium channel subunit, or both.

1           17.    A eukaryotic cell transiently or stably transformed with a heterologous DNA  
2 fragment comprising a sequence of nucleotides that encodes an  $\alpha_1$  subunit of a mammalian T-  
3 type calcium channel, wherein the cell expresses the  $\alpha_1$  subunit of a mammalian T-type  
4 calcium channel.

1           18.    The cell of claim 17 , wherein the cell is further transformed with and  
2 expresses an  $\alpha_2\delta$  or a  $\beta$  calcium channel subunit, or both.

1           19.    A method for the production of an  $\alpha_1$  subunit of a mammalian T-type calcium  
2 channel comprising, culturing a cell transiently or stably transformed or transfected with DNA  
3 encoding the calcium channel subunit under conditions such that the DNA is expressed and  
4 the  $\alpha_1$  subunit is produced.

1           20. A process for producing a eukaryotic cell that is transiently or stably transformed  
2 and expresses a mammalian T-type calcium channel, comprising the step of introducing RNA  
3 or DNA encoding an  $\alpha_1$  subunit of a mammalian T-type calcium channel.

1           21. A method of identifying compounds capable of acting as agonists or antagonists  
2 for T-type mammalian calcium channels, comprising contacting a recombinant cell expressing  
3 a heterologous  $\alpha_1$  subunit of a mammalian T-type calcium channel, with an agent to be tested,  
4 and evaluating the interaction, if any, between the agent to be tested and the calcium channel.

1           22.    An isolated DNA fragment having the sequence given by SEQ ID No. 19, 27  
2           or 29.

1           23.    A method for mapping the distribution of T-type calcium channels within a  
2           tissue sample comprising the steps of exposing the tissue to a reagent comprising a directly-  
3           or indirectly-detectable label coupled to a DNA fragment comprising a sequence selected  
4           from among those sequences given by SEQ ID Nos. 13-17, 19, 23, 25, 27, 29, 30, 32 and 34,  
5           and detecting reagent that has bound to the tissue.

1           24.    A DNA fragment comprising a sequence of oligonucleotide that encodes a  
2           calcium channel, wherein the sequence of nucleotides is selected from sequences of  
3           nucleotides encoding a protein including the sequence of amino acids set forth in SEQ ID. No.  
4           18, 20, 24, 26, 28, 31, 33, or 35, or a nucleotide sequence obtainable by subcloning a PCR  
5           product of SEQ ID Nos: 13, 14, 15, or 16, labeling it by random hexamer priming, using the  
6           primer to screen a commercial human brain cDNA library to produce partial sequence clones  
7           containing overlapping cDNAs and ligating cDNAs obtained to produce full length cDNA  
8           encoding a calcium channel protein.



ABSTRACT OF THE DISCLOSURE

1                   Sequences and partial sequences for three types of mammalian (human and rat  
2 sequences identified) T-type calcium channel subunits which we have labeled as the  $\alpha_{1G}$ ,  $\alpha_{1H}$   
3 and  $\alpha_{1I}$  subunits are provided. Knowledge of the sequence of these calcium channel permits  
4 the localization and recovery of the complete sequence from human cells, and the  
5 development of cell lines which express the novel calcium channels of the invention. These  
6 cells may be used for identifying compounds capable of acting as agonists or antagonists to  
7 the calcium channels.

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant: Snutch et al.  
Serial No.: to be assigned  
Filed: herewith  
For: NOVEL HUMAN CALCIUM CHANNELS AND RELATED PROBES, CELL  
LINES AND METHODS

Statement Regarding Sequence Listing

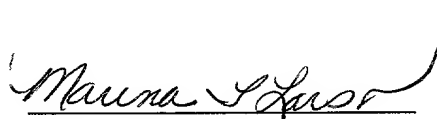
Asst. Commissioner for Patents

Washington, D.C. 20231

Sir:

The undersigned hereby certifies that the paper copy of the sequence listing and the machine readable diskette filed herewith contain the same information.

Respectfully submitted,



Marina T. Larson  
PTO Reg. No. 32,038  
Attorney for Applicant  
(970) 668-2050

1/4

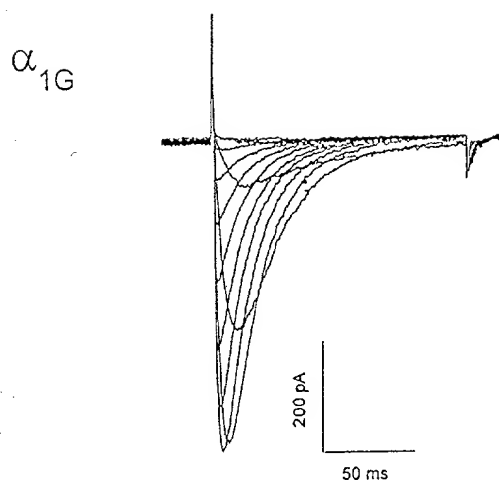


Fig. 1A

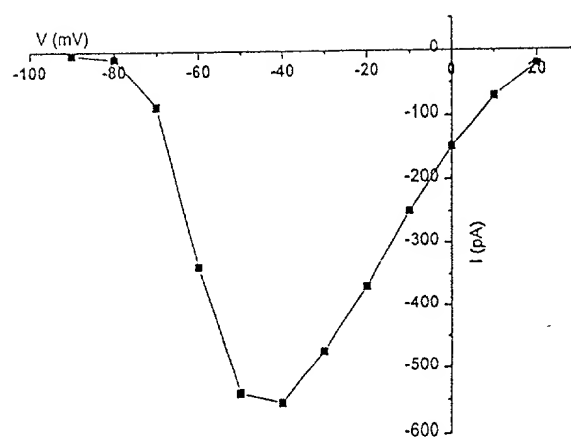


Fig. 1B

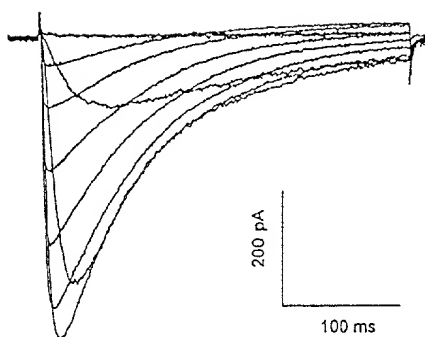
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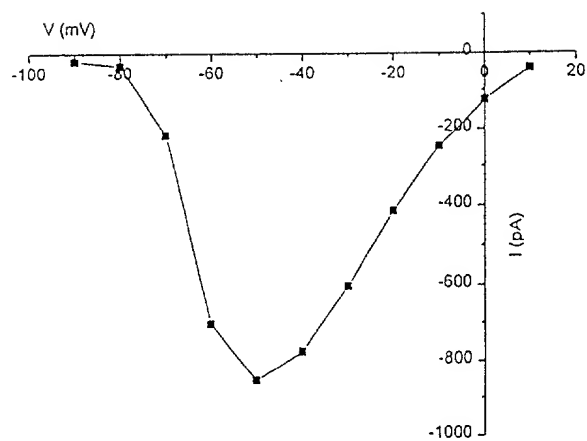


Fig. 2 B

## Steady-state inactivation

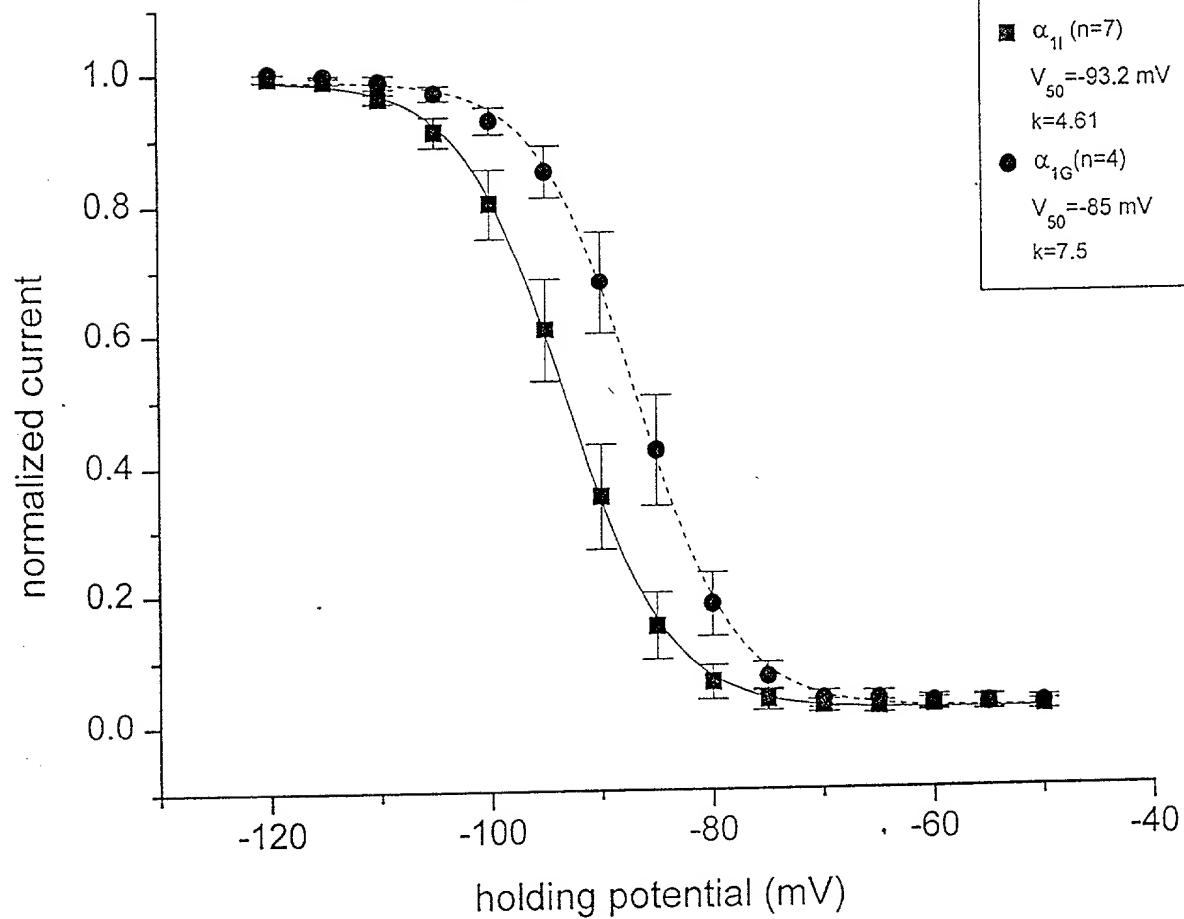


Fig. 3

Deactivation

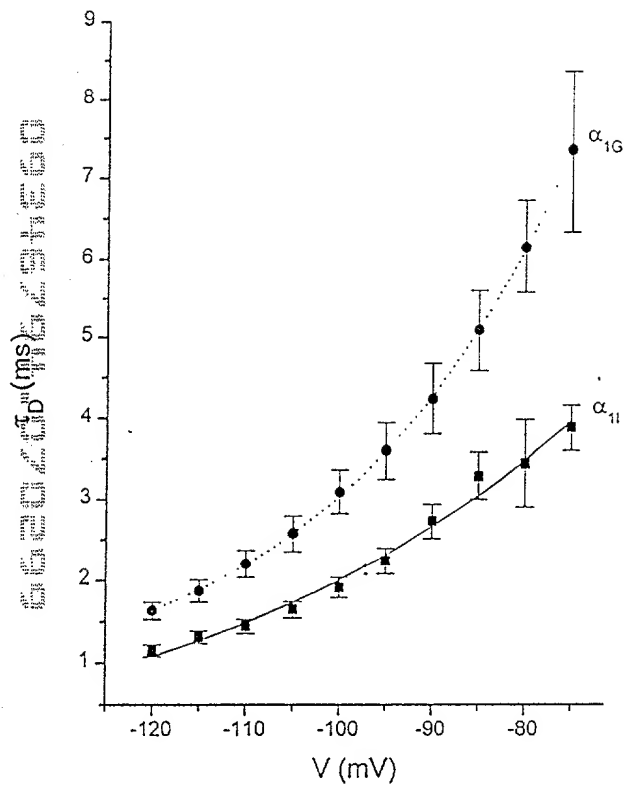


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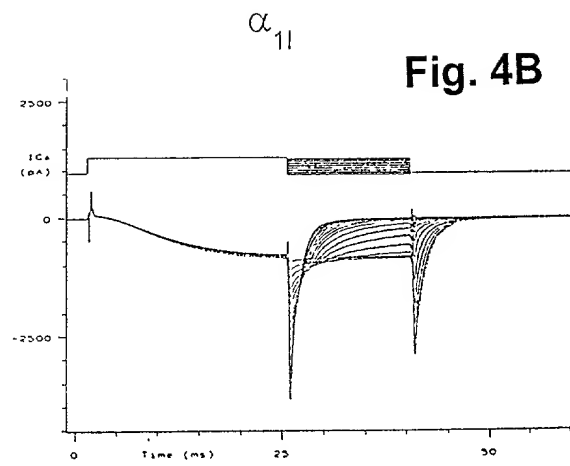


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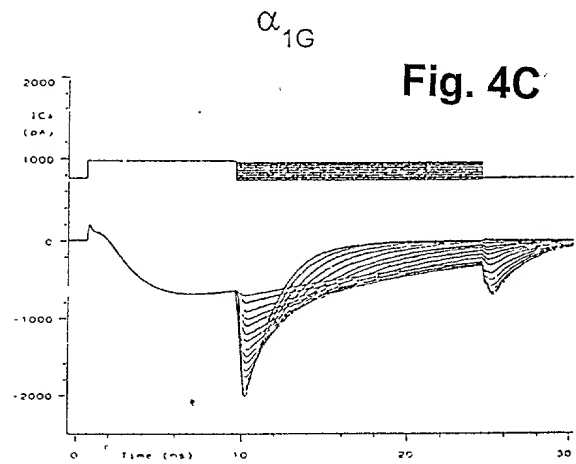


Fig. 4C

# SEQUENCE LISTING

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Baillie, David L.

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LINES AND METHODS

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CC-BY-NC-SA

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Asn Ile Asn Leu Ser Ala Ile Arg Thr Val Arg Val Leu Arg Pro Leu		
210	215	220
Lys Ala Ile Asn Arg Val Pro Ser Met Arg Ile Leu Val Asn Leu Leu		
225	230	235 240
Leu Asp Thr Leu Pro Met Leu Gly Asn Val Leu Leu Leu Cys Phe Phe		
245	250	255
Val Phe Phe Ile Phe Gly Ile Ile Gly Val Gln Leu Trp Ala Gly Leu		
260	265	270
Leu Arg Asn Arg Cys Phe Leu Glu Glu Asn Phe Thr Ile Gln Gly Asp		
275	280	285
Val Ala Leu Pro Pro Tyr Tyr Gln Pro Glu Glu Asp Asp Glu Met Pro		

290

295

300

Phe Ile Cys Ser Leu Ser Gly Asp Asn Gly Ile Met Gly Cys His Glu  
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Ile Pro Pro Leu Lys Glu Gln Gly Arg Glu Cys Cys Leu Ser Lys Asp  
325 330 335

Asp Val Tyr Asp Phe Gly Ala Gly Arg Gln Asp Leu Asn Ala Ser Gly  
340 345 350

Leu Cys Val Asn Trp Asn Arg Tyr Tyr Asn Val Cys Arg Thr Gly Ser  
355 360 365

Ala Asn Pro His Lys Gly Ala Ile Asn Phe Asp Asn Ile Gly Tyr Ala  
370 375 380

Trp Ile Val Ile Phe Gln Val Ile Thr Leu Glu Gly Trp Val Glu Ile  
385 390 395 400

Met Tyr Tyr Val Met Asp Ala His Ser Phe Tyr Asn Phe Ile Tyr Phe  
405 410 415

Ile Leu Leu Ile Ile Ser Glu Leu Ile His Leu Val Met Pro Asp Cys  
420 425 430

Ser Phe Ser Thr Ala Gln Ser Pro Lys Cys Gln Gly Asp Ser Leu Pro  
435 440 445

Gly Val Ala Ala Glu Ser Leu Leu Leu Arg Asp Ser Ser Ser Ser Val  
450 455 460

Ile Thr Asp Glu Ala Ala Ala Met Glu Asn Leu Leu Ala Gly Thr Ser  
465 470 475 480

Lys Gly Asp Glu Ser Tyr Leu Leu Arg Leu Ala Gly Ser Gln Val His  
485 490 495

Ser Gln Ala Gln Gln Met Leu Gly Arg Gly Leu Gly Pro Glu Ser Leu  
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Glu Thr Gly Glu Glu Pro His Ser Trp Ser Pro Arg Ala Thr Arg Arg  
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Trp Asp Pro Gln Cys Gln Pro Gly Gln Pro Leu Pro Leu His Phe Met  
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Gln Ala Gln Val Gly Ser Phe Phe Met Ile Asn Leu Cys Leu Val Val

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Ile Ala Thr Gln Phe Ser Glu Thr Lys Gln Arg Glu His Arg Leu Met			
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Leu Glu Gln Arg Gln Arg Tyr Leu Ser Ser Ser Thr Val Ala Ser Tyr			
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Ala Glu Pro Gly Asp Cys Tyr Glu Glu Ile Phe Gln Tyr Val Cys His			
595	600	605	
Ile Leu Arg Lys Ala Lys Arg Arg Ala Leu Gly Leu Tyr Gln Ala Leu			
610	615	620	
Gln Ser Arg Arg Gln Ala Leu Gly Pro Glu Ala Pro Ala Pro Ala Lys			
625	630	635	640
Pro Gly Pro His Ala Lys Glu Pro Arg His Tyr Pro Leu Thr Val Trp			
645	650	655	
Glu Ser Ile Leu Gly Arg Gln Ala Glu Glu Cys Thr Leu Arg Ala Ala			
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Ala His Pro Ser Ser Gly Ala Ser His Pro Gly Val Gly Ser Glu Glu			
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Ala Pro Glu Leu Cys Pro Gln His Ser Pro Leu Asp Ala Thr Pro His			
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Cys Pro Cys Cys Gln His Glu Asp Gly Arg Arg Pro Ser Gly Leu Gly			
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Ser Thr Asp Ser Gly Gln Glu Gly Ser Gly Ser Gly Ser Ser Ala Gly			
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Gly Glu Asp Glu Ala Asp Gly Asp Gly Ala Arg Ser Ser Glu Asp Gly			
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Ala Ser Ser Glu Leu Gly Lys Glu Glu Glu Glu Glu Glu Gln Ala Asp			
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Gly Ala Val Trp Leu Cys Gly Asp Val Trp Arg Glu Thr Arg Ala Lys			
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Leu Arg Gly Ile Val Asp Ser Lys Tyr Phe Asn Arg Gly Ile Met Met			



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Asp Leu Cys Met Thr Leu Lys Ala Pro Cys Leu Cys His Asn Val Pro		
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Ser Pro Gly Gln Gly Val Leu Ser His Pro Val Thr Pro Pro His Thr		
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Ala Pro Trp Arg Met Glu Thr Gly Lys Gln Gly His Gly Cys Glu Glu		
885	890	895
Gly Pro Gly Gln Arg Ser Ser Asp Met Phe Ala Leu Glu Met Ile Leu		
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Lys Leu Ala Ala Phe Gly Leu Phe Asp Tyr Leu Arg Asn Pro Tyr Asn		
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Ile Phe Asp Ser Ile Ile Val Ile Ile Ser Ile Trp Glu Ile Val Gly		
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Gln Ala Asp Gly Gly Leu Ser Val Leu Arg Thr Phe Arg Leu Leu Arg		
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Val Leu Lys Leu Val Arg Phe Met Pro Ala Leu Arg Arg Gln Leu Val		
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Val Leu Met Lys Thr Met Asp Asn Val Ala Thr Phe Cys Met Leu Leu		
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Met Leu Phe Ile Phe Ile Phe Ser Ile Leu Gly Met His Ile Phe Gly		
995	1000	1005
Cys Lys Phe Ser Leu Arg Thr Asp Thr Gly Asp Thr Val Pro Asp Arg		
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Lys Asn Phe Asp Ser Leu Leu Trp Ala Ile Val Thr Val Phe Gln Ile		
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Leu Thr Gln Glu Asp Trp Asn Val Val Leu Tyr Asn Gly Met Ala Ser		
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Thr Ser Pro Trp Ala Ser Leu Tyr Phe Val Ala Leu Met Thr Phe Gly		

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Asn Tyr Val Leu Phe Asn Leu Leu Val Ala Ile Leu Val Glu Gly Phe		
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Gln Ala Glu Val Thr Val Val Leu Ala Glu Glu Ala Pro Pro Gln Gly		
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Leu Arg Lys Thr Gly Arg Gly Arg Gly Gly Leu Asp Gly Gly Gly Leu		
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Gln Phe Lys Leu Leu Ala Gly Asn Leu Ser Leu Lys Glu Gly Val Ala		
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Asp Glu Val Gly Asp Ala Asn Arg Ser Tyr Ser Asp Glu Asp Gln Ser		
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Ser Ser Asn Ile Glu Glu Phe Asp Lys Leu Gln Glu Gly Leu Asp Ser		
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Ser Gly Asp Pro Lys Leu Cys Pro Ile Pro Met Thr Pro Asn Gly His		
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Leu Asp Pro Ser Leu Pro Leu Gly Gly His Leu Gly Pro Ala Gly Ala		
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Ala Gly Pro Ala Pro Arg Leu Ser Leu Gln Pro Asp Pro Met Leu Val		
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Ala Leu Gly Ser Arg Lys Ser Ser Val Met Ser Leu Gly Arg Met Ser		
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Tyr Asp Gln Arg Ser Leu Val Gly Gly Leu Arg Ala Thr Ala Gly Val		
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Gln Ala Ala Phe Gly His Leu Val Pro Gln Pro Trp Val Cys Leu Trp		
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Gly Ala Asp Pro Asn Gly Asn Ser Phe Gln Ser Ser Ser Arg Ser Ser		
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Tyr Tyr Gly Pro Trp Gly Arg Ser Ala Ala Trp Ala Ser Arg Arg Ser		
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Ser Trp Asn Ser Leu Lys His Lys Pro Pro Ser Ala Glu His Glu Ser		
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His His Val His His Gly Pro His Leu Ala His Arg His Arg His His		
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Arg Arg Thr Leu Ser Leu Asp Asn Arg Asp Ser Val Asp Leu Ala Glu		
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Leu Val Pro Ala Val Gly Ala His Pro Arg Ala Ala Trp Arg Ala Ala		
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Gly Pro Ala Pro Gly His Glu Asp Cys Asn Gly Arg Met Pro Ser Ile		
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Ala Lys Asp Val Phe Thr Lys Met Gly Asp Arg Gly Asp Arg Gly Glu		
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Asp Glu Glu Glu Ile Asp Tyr Val Ser Gly Gly Gly Ala Glu Gly Asp		
1425	1430	1435 1440
Leu Thr Leu Cys Phe Arg Val Arg Lys Met Ile Asp Val Tyr Lys Pro		
1445	1450	1455
Asp Trp Cys Glu Val Arg Glu Asp Trp Ser Val Tyr Leu Phe Ser Pro		
1460	1465	1470
Glu Asn Arg Leu Arg Asp Leu Gly Trp Val Ser Leu Glu Cys Gln Gly		
1475	1480	1485
Lys Val Gly Asp Leu Val Val Trp Val Tyr Gly Gln Arg Arg Gln Arg		
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Gln Thr Ile Ile Ala His Lys Leu Phe Asp Tyr Val Val Leu Ala Phe		
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Ile Phe Leu Asn Cys Ile Thr Ile Ala Leu Glu Arg Pro Gln Ile Glu		
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Ala Gly Ser Thr Glu Arg Ile Phe Leu Thr Val Ser Asn Tyr Ile Phe		
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Thr Ala Ile Phe Val Gly Glu Met Thr Leu Lys Val Val Ser Leu Gly		
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1570

1575

1580

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Gly Val Gln Leu Phe Lys Gly Lys Phe Tyr His Cys Leu Gly Val Asp  
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Asn Pro Trp Met Leu Leu Tyr Phe Ile Ser Phe Leu Leu Ile Val Ser  
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Phe Phe Val Leu Asn Met Phe Val Gly Val Val Val Glu Asn Phe His  
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Lys Cys Arg Gln His Gln Glu Ala Glu Glu Ala Arg Arg Arg Glu Glu  
 1780 1785 1790

Lys Arg Leu Arg Arg Leu Glu Lys Lys Arg Arg Lys Ala Gln Arg Leu  
 1795 1800 1805

Pro Tyr Tyr Ala Thr Tyr Cys His Thr Arg Leu Leu Ile His Ser Met  
 1810 1815 1820

Cys Thr Ser His Tyr Leu Asp Ile Phe Ile Thr Phe Ile Ile Cys Leu

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<213> HUMAN

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20 25 30  
Gly Val Gln Leu Trp Ala Gly Leu Leu Arg Asn Arg Cys Phe Leu Glu  
35 40 45  
Glu Asn Phe Thr Ile Gln Gly Asp Val Ala Leu Pro Pro Tyr Tyr Gln  
50 55 60  
Pro Glu Glu Asp Asp Glu Met Pro Phe Ile Cys Ser Leu Ser Gly Asp

65	70	75	80
Asn Gly Ile Met Gly Cys His Glu Ile Pro Pro Leu Lys Glu Gln Gly			
85	90	95	
Arg Glu Cys Cys Leu Ser Lys Asp Asp Val Tyr Asp Phe Gly Ala Gly			
100	105	110	
Arg Gln Asp Leu Asn Ala Ser Gly Leu Cys Val Asn Trp Asn Arg Tyr			
115	120	125	
Tyr Asn Val Cys Arg Thr Gly Ser Ala Asn Pro His Lys Gly Ala Ile			
130	135	140	
Ser Phe Asp Asn Ile Gly Tyr Ala Trp Ile Val Ile Phe Gln Val Ile			
145	150	155	160
Thr Leu Glu Gly Trp Val Ala Ile Met Tyr Tyr Val Met Asp Ala Leu			
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<212> DNA

<213> rat

<220>

<223> rat alpha-I partial sequence

<400> 21

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<211> 189

<212> PRT

<213> rat

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<223> rat alpha-I partial sequence

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20 25 30

Gly Val Gln Leu Trp Ala Gly Leu Leu Arg Asn Arg Cys Phe Leu Glu  
35 40 45

Glu Asn Phe Thr Ile Gln Gly Asp Val Ala Leu Pro Pro Tyr Tyr Gln  
50 55 60

Pro Glu Glu Asp Asp Glu Met Pro Phe Ile Cys Ser Leu Thr Gly Asp  
65 70 75 80

Asn Gly Ile Met Gly Cys His Glu Ile Pro Pro Leu Lys Glu Gln Gly  
85 90 95

Arg Glu Cys Cys Leu Ser Lys Asp Asp Val Tyr Asp Phe Gly Ala Gly  
100 105 110

Arg Gln Asp Leu Asn Ala Ser Gly Leu Cys Val Asn Trp Asn Arg Tyr  
115 120 125

Tyr Asn Val Cys Arg Thr Gly Asn Ala Asn Pro His Lys Gly Ala Ile  
130 135 140

Asn Phe Asp Asn Ile Gly Tyr Ala Trp Ile Val Ile Phe Gln Val Ile  
145 150 155 160

Thr Leu Glu Gly Trp Val Glu Ile Met Tyr Tyr Val Met Asp Ala His  
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<213> rat

<400> 26

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 35 40 45  
 Leu Ala Pro Glu Ser Pro Gly Thr Glu Cys Gly Ala Asp Leu Gly Ala  
 50 55 60  
 Asp Glu Glu Gln Pro Val Pro Tyr Pro Ala Leu Ala Ala Thr Val Phe  
 65 70 75 80  
 Phe Cys Leu Gly Gln Thr Thr Arg Pro Arg Ser Trp Cys Leu Arg Leu  
 85 90 95  
 Val Cys Asn Pro Trp Phe Glu His Val Ser Met Leu Val Ile Met Leu  
 100 105 110  
 Asn Cys Val Thr Leu Gly Met Phe Arg Pro Cys Glu Asp Val Glu Cys  
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 Arg Ser Glu Arg Cys Ser Ile Leu Glu Ala Phe Asp Asp Phe Ile Phe  
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 Phe Gly Gln Lys Cys Tyr Leu Gly Asp Thr Trp Asn Arg Leu Asp Phe  
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 Phe Ile Val Met Ala Gly Met Met Glu Tyr Ser Leu Asp Gly His Lys  
 180 185 190  
 Val Ser Leu Ser Ala Ile Arg Thr Val Arg Val Leu Arg Pro Leu Arg  
 195 200 205  
 Ala Ile Asn Arg Val Pro Ser Met Arg Ile Leu Val Thr Leu Leu Leu  
 210 215 220  
 Asp Thr Leu Pro Met Leu Gly Asn Val Leu Leu Leu Cys Phe Phe Val  
 225 230 235 240  
 Phe Phe Ile Phe Gly Ile Val Gly Val Gln Leu Trp Ala Gly Leu Leu  
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 Arg Asn Arg Cys Phe Leu Asp Ser Ala Phe Val Arg Asn Asn Asn Leu  
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Thr	Phe	Leu	Arg	Pro	Tyr	Tyr	Gln	Thr	Glu	Glu	Gly	Glu	Glu	Asn	Pro	275	280	285	
Phe	Ile	Cys	Ser	Ser	Arg	Arg	Asp	Asn	Gly	Met	Gln	Lys	Cys	Ser	His	290	295	300	
Ile	Pro	Ser	Arg	Arg	Glu	Leu	Arg	Val	Gln	Cys	Thr	Leu	Gly	Trp	Glu	305	310	315	320
Ala	Tyr	Gly	Gln	Pro	Gln	Ala	Glu	Asp	Gly	Gly	Ala	Gly	Arg	Asn	Ala	325	330	335	
Cys	Ile	Asn	Trp	Asn	Gln	Tyr	Tyr	Asn	Val	Cys	Arg	Ser	Gly	Glu	Phe	340	345	350	
Asn	Pro	His	Asn	Gly	Ala	Ile	Asn	Phe	Asp	Asn	Ile	Gly	Tyr	Ala	Trp	355	360	365	
Ile	Ala	Ile	Phe	Gln	Val	Ile	Thr	Leu	Glu	Gly	Trp	Val	Asp	Ile	Met	370	375	380	
Tyr	Tyr	Val	Met	Asp	Ala	His	Ser	Phe	Tyr	Asn	Phe	Ile	Tyr	Phe	Ile	385	390	395	400
Leu	Leu	Ile	Ile	Met	Gly	Ser	Phe	Phe	Met	Ile	Asn	Leu	Cys	Leu	Val	405	410	415	
Val	Ile	Ala	Thr	Gln	Phe	Ser	Glu	Thr	Lys	Gln	Arg	Glu	Asn	Gln	Leu	420	425	430	
Met	Arg	Glu	Gln	Arg	Ala	Arg	Tyr	Leu	Ser	Asn	Asp	Ser	Thr	Leu	Ala	435	440	445	
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Arg	Trp	Gln	Ser	Arg	Trp	Arg	Lys	Lys	Val	Asp	Pro	Ser	Ser	Thr	Val	485	490	495	
His	Gly	Gln	Gly	Pro	Gly	Arg	Arg	Pro	Arg	Arg	Ala	Gly	Arg	Arg	Thr	500	505	510	
Ala	Ser	Val	His	His	Leu	Val	Tyr	His	His	His	His	His	His	His	His	515	520	525	



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 545 550 555 560  
 Pro Ser Pro Gly His Gly Pro Pro Asp Ser Glu Ser Val His Ser Ile  
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 Tyr His Ala Asp Cys His Val Glu Gly Pro Gln Glu Arg Ala Arg Val  
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 Ser Ser His Leu Ser Gly Leu Ser Val Pro Cys Pro Leu Pro Ser Pro  
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 Gln Ala Gly Thr Leu Thr Cys Glu Leu Lys Ser Cys Pro Tyr Cys Ala  
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 Ser Ala Leu Glu Asp Pro Glu Phe Glu Phe Ser Gly Ser Glu Ser Gly  
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 Asp Ser Asp Ala His Gly Val Tyr Glu Phe Thr Gln Asp Val Arg His  
 725 730 735  
 Gly Asp Cys Arg Asp Pro Val Gln Gln Pro His Glu Val Gly Thr Pro  
 740 745 750  
 Gly His Ser Asn Glu Arg Arg Arg Thr Pro Leu Arg Lys Ala Ser Gln  
 755 760 765  
 Pro Gly Gly Ile Gly His Leu Trp Ala Ser Phe Ser Gly Lys Leu Arg  
 770 775 780

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Glu	Leu	Thr	Asn	Ala	Leu	Glu	Ile	Ser	Asn	Ile	Val	Phe	Thr	Ser	Met				
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Phe	Ala	Leu	Glu	Met	Leu	Leu	Lys	Leu	Leu	Ala	Cys	Gly	Pro	Leu	Gly				
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Tyr	Ile	Arg	Asn	Pro	Tyr	Asn	Ile	Phe	Asp	Gly	Ile	Val	Val	Val	Ile				
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Ser	Val	Trp	Glu	Ile	Val	Gly	Gln	Ala	Asp	Gly	Gly	Gln	Ser	Val	Leu				
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Arg	Thr	Phe	Arg	Leu	Leu	Arg	Val	Leu	Lys	Leu	Val	Arg	Phe	Leu	Pro				
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Ala	Thr	Phe	Cys	Met	Leu	Leu	Met	Leu	Phe	Ile	Phe	Ile	Phe	Ser	Ile				
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Ala	Ile	Leu	Val	Glu	Gly	Phe	Gln	Ala	Glu	Gly	Asp	Ala	Thr	Arg	Ser				
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 Pro Asn Leu Asp Val Ala His Ala Leu Leu Asp Ser Arg Arg Ser Ser  
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 Ser Gly Ser Val Asp Pro Gln Leu Gly Asp Gln Lys Ser Leu Ala Ser  
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Val Phe Glu Ala Ala Leu Lys Leu Val Ala Phe Gly Phe Arg Arg Phe  
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Thr Val Val Gln Ala Leu Pro Gln Val Gly Asn Leu Gly Leu Leu Phe  
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Met Leu Leu Phe Phe Ile Tyr Ala Ala Leu Gly Val Glu Leu Phe Gly  
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Arg Leu Glu Cys Ser Glu Asp Asn Pro Cys Glu Gly Leu Ser Arg His  
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Ala Thr Phe Thr Asn Phe Gly Met Ala Phe Leu Thr Leu Phe Arg Val  
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Ser Thr Gly Asp Asn Trp Asn Gly Ile Met Lys Asp Thr Leu Arg Glu  
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Cys Thr Arg Glu Asp Lys His Cys Leu Ser Tyr Leu Pro Ala Leu Ser  
 1825 1830 1835 1840

Pro Val Tyr Phe Val Thr Phe Met Leu Val Ala Gln Phe Val Leu Val  
 1845 1850 1855

Asn Val Val Val Ala Val Leu Met Lys His Leu Glu Glu Ser Asn Lys  
 1860 1865 1870

Glu Ala Arg Glu Asp Ala Glu Met Asp Ala Glu Ile Glu Leu Glu Met  
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Gly Thr Gln Pro Asp Thr Pro Asn Leu Leu Val Val Arg Lys Val Ser  
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Phe Leu Asp Lys Pro Gly Arg Pro Asp Ala Gln Arg Trp Ser Ser Val  
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Glu Leu Asp Asn Gly Glu Ser His Leu Glu Ser Gly Glu Val Arg Gly  
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Arg Ala Ser Glu Leu Glu Pro Ala Leu Gly Ser Arg Arg Lys Lys Lys  
                          2180                      2185                      2190

Met Ser Pro Pro Cys Ile Ser Ile Glu Pro Pro Thr Lys Asp Glu Gly  
                          2195                      2200                      2205

Ser Ser Arg Pro Pro Ala Ala Glu Gly Gly Asn Thr Thr Leu Arg Arg  
                          2210                      2215                      2220

Arg Thr Pro Ser Cys Glu Ala Ala Leu His Arg Asp Cys Pro Glu Pro  
 2225                      2230                      2235                      2240

Thr Glu Gly Pro Gly Thr Gly Gly Asp Pro Val Ala Lys Gly Glu Arg  
                          2245                      2250                      2255

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His Pro Asp Leu Ala Pro Val Ala Phe Phe Cys Leu Arg Gln Thr Thr
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Ser Pro Arg Asn Trp Cys Ile Lys Met Val Cys Asn Pro Trp Phe Glu
      65              70              75              80

Cys Val Ser Met Leu Val Ile Leu Leu Asn Cys Val Thr Leu Gly Met
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Tyr Gln Pro Cys Asp Asp Met Glu Cys Leu Ser Asp Arg Cys Lys Ile
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Val Leu Lys Met Val Ala Leu Gly Ile Phe Gly Lys Lys Cys Tyr Leu

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Tyr Asn Val Cys Arg Thr Gly Asn Ala Asn Pro His Lys Gly Ala Ile					
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Thr Leu Val Gln	Pro Ile Ser Ala Ile	Leu Ala Ser Tyr	Pro Ser Ser			
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Cys Gly Asp Val	Trp Arg Glu Thr Arg	Lys Lys Leu Arg	Gly Ile Val			
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Asp Ser Lys Tyr	Phe Asn Arg Gly Ile	Met Met Ala Ile	Leu Val Asn			
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Gln Leu Val Val Leu Met Lys Thr Met Asp Asn Val Ala Thr Phe Cys		
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Met Leu Leu Met Leu Phe Ile Phe Ile Phe Ser Ile Leu Gly Ile Asp		
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Glu Gly Leu Pro Tyr Pro Ala Leu Ala Pro Val Val Phe Phe Tyr Leu  
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Ser Gln Asp Ser Arg Pro Arg Ser Trp Cys Leu Arg Thr Val Cys Asn  
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Pro Trp Phe Glu Arg Ile Ser Met Leu Val Ile Leu Leu Asn Cys Val  
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Thr Leu Gly Met Phe Arg Pro Cys Glu Asp Ile Ala Cys Asp Ser Gln  
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Arg Cys Arg Ile Leu Gln Ala Phe Asp Asp Phe Ile Phe Ala Phe Phe  
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Ala Val Glu Met Val Val Lys Met Val Ala Leu Gly Ile Phe Gly Lys  
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Lys Cys Tyr Leu Gly Asp Thr Trp Asn Arg Leu Asp Phe Phe Ile Val  
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Ile Ala Gly Met Leu Glu Tyr Ser Leu Asp Leu Gln Asn Val Ser Phe  
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Ser Ala Val Arg Thr Val Arg Val Leu Arg Pro Leu Arg Ala Ile Asn  
 180 185 190

Arg Val Pro Ser Met Arg Ile Leu Val Thr Leu Leu Leu Asp Thr Leu  
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Pro Met Leu Gly Asn Val Leu Leu Leu Cys Phe Phe Val Phe Phe Ile  
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Cys Phe Leu Pro Glu Asn Phe Ser Leu Pro Leu Ser Val Asp Leu Glu  
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Arg Tyr Tyr Gln Thr Glu Asn Glu Asp Glu Ser Pro Phe Ile Cys Ser  
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Gln Pro Arg Glu Asn Gly Met Arg Ser Cys Arg Ser Val Pro Thr Leu  
 275 280 285

Arg Gly Asp Gly Gly Gly Gly Pro Pro Cys Gly Leu Asp Tyr Glu Ala  
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Tyr Asn Ser Ser Ser Asn Thr Thr Cys Val Asn Trp Asn Gln Tyr Tyr  
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Thr Asn Cys Ser Ala Gly Glu His Asn Pro Phe Lys Gly Ala Ile Asn  
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Phe Asp Asn Ile Gly Tyr Ala Trp Ile Ala Ile Phe Gln Val Ile Thr  
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Leu Glu Gly Trp Val Asp Ile Met Tyr Phe Val Met Asp Ala His Ser  
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Phe Tyr Asn Phe Ile Tyr Phe Ile Leu Leu Ile Ile Val Gly Ser Phe  
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Thr Lys Gln Arg Glu Ser Gln Leu Met Arg Glu Gln Arg Val Arg Phe  
 405 410 415

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Tyr Glu Glu Leu Leu Lys Tyr Leu Val Tyr Ile Leu Arg Lys Ala Ala  
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Arg Arg Leu Ala Gln Val Ser Arg Ala Ala Gly Val Arg Val Gly Leu  
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Ser Cys Ser Arg Ser His Arg Arg Leu Ser Val His His Leu Val His  
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His His His His His His His His Tyr His Leu Gly Asn Gly Thr Leu  
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Cys His Leu Glu Pro Val Arg Cys Gln Ala Pro Pro Pro Arg Ser Pro  
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Val His Thr Ser Pro Pro Pro Glu Thr Leu Lys Glu Lys Ala Leu Val  
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Glu Val Ala Ala Ser Ser Gly Pro Pro Thr Leu Thr Ser Leu Asn Ile  
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Ile Ala Ile Phe Gln Val Ile Thr Leu Glu Gly Trp Val Asp Ile Met					
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Tyr Tyr Val Met Asp Ala His Ser Phe Tyr Asn Phe Ile Tyr Phe Ile					
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Leu Leu Ile Ile Val Gly Ser Phe Phe Met Ile Asn Leu Cys Leu Val					
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Val Ile Ala Thr Gln Phe Ser Glu Thr Lys Gln Arg Glu Ser Gln Leu					

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Val Cys Glu Val Ala Ala Asp Glu Gly Pro Pro Arg Ala Ala Pro Leu  
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His Thr Pro His Ala His His Ile His His Gly Pro His Leu Ala His  
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Arg His Arg His His Arg Arg Thr Leu Ser Leu Asp Asn Arg Asp Ser  
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Val Asp Leu Ala Glu Leu Val Pro Ala Val Gly Ala His Pro Arg Ala  
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Arg Val Arg Lys Met Ile Asp Val Tyr Lys Pro Asp Trp Cys Glu Val  
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Arg Glu Asp Trp Ser Val Tyr Leu Phe Ser Pro Glu Asn Arg Phe Arg  
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Gln Ile Glu Ala Gly Ser Thr Glu Arg Ile Phe Leu Thr Val Ser Asn  
245 250 255

Tyr Ile Phe Thr Ala Ile Phe Val Gly Glu Met Thr Leu Lys Val Val  
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Asn Val Leu Asp Gly Phe Leu Val Phe Val Ser Ile Ile Asp Ile Val  
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Val Ser Leu Ala Ser Ala Gly Gly Ala Lys Ile Leu Gly Val Leu Arg  
305 310 315 320

Val Leu Arg Leu Leu Arg Thr Leu Arg Pro Leu Arg Val Ile Ser Arg  
325 330 335

Ala Pro Gly Leu Lys Leu Val Val Glu Thr Leu Ile Ser Ser Leu Lys  
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Pro Ile Gly Asn Ile Val Leu  
355

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FILE NO. NMED.P-001-2**COMBINED DECLARATION  
AND POWER OF ATTORNEY**

As a below named inventor, I hereby declare that:

My citizenship, residence and post office address are as listed below next to my name.

I believe I am the original, first and [ ] sole/[x] joint inventor of the subject matter which is claimed and for which a patent is sought on the invention entitled: Novel Human Calcium Channels and Related Probes, Cell Lines and Methods  
the specification of which

(a) [X] is attached hereto.

(b) [ ] was filed on \_\_\_\_\_ as Application Serial No. \_\_\_\_\_ and was amended on \_\_\_\_\_.

(c) [ ] was described and claimed in International Application No. \_\_\_\_\_ filed on \_\_\_\_\_ and amended on \_\_\_\_\_.

**Acknowledgment of Duty of Disclosure**

I hereby state that I have reviewed and understood the content of the above identified specification, including the claims, as amended by any amendment referred to above. I acknowledge the duty to disclose information which is material to the patentability of the subject matter claimed in this application in accordance with Title 37, Code of Federal Regulations § 1.56(a).

**Continuation-In-Part Application**

I hereby claim the benefit under Title 35, United States Code, § 120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code § 112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal regulations, § 1.56(a) which occurred between the filing date of the prior application and the national or PCT international filing date of this application:

<u>09/030,482</u>	<u>February 25, 1999</u>	<u>pending</u>
(Application Serial No.)	(Filing Date)	(Status)(patented,pending,abandoned)

_____	_____	_____
(Application Serial No.)	(Filing Date)	(Status)(patented,pending,abandoned)

**Power of Attorney**

I hereby appoint Carl Oppedahl, PTO Reg. NO. 32,746 and Marina T. Larson, PTO Reg. No. 32,038 of the firm of OPPEDAHL & LARSON LLP, whose address is PO Box 5270, 611 main Street, Frisco, CO 80443-5270 as attorneys to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith.

SEND CORRESPONDENCE TO:  
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PO BOX 5270  
FRISCO, CO 80443-5270

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OPPEDAHL & LARSON  
(970) 668-2050

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FILE NO. NMED.P-001-2**Claim for Priority**

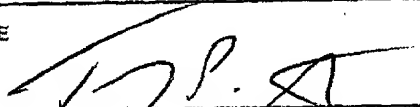
I hereby claim priority under Title 35, United States Code, § 119 of any foreign application(s) for patent or inventor's certificate listed below and have identified any foreign applications for patent or inventor's certificate having a filing date before that of the application on which priority is claimed.

EARLIEST FOREIGN APPLICATION(S), FILED WITHIN TWELVE MONTHS (6 MONTHS FOR DESIGN) PRIOR TO SAID APPLICATION				
COUNTRY	APPLICATION NO.	DATE OF FILING (day/month/year)	DATE OF ISSUE (day/month/year)	PRIORITY CLAIMED
				YES[ ] NO[ ]
				YES[ ] NO[ ]
				YES[ ] NO[ ]

FOREIGN APPLICATION(S), IF ANY, FILED MORE THAN 12 MONTHS (6 MONTHS FOR DESIGN) PRIOR TO SAID APPLICATION				
COUNTRY	APPLICATION NO.	DATE OF FILING (day/month/year)	DATE OF ISSUE (day/month/year)	

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements are made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

NAME OF SOLE OR FIRST INVENTOR	LAST NAME <b>SNUTCH</b>	FIRST NAME <b>TERRY</b>	MIDDLE NAME <b>P.</b>
RESIDENCE & CITIZENSHIP	CITY OF RESIDENCE <b>VANCOUVER</b>	STATE OR COUNTRY OF RESIDENCE <b>CANADA</b>	COUNTRY OF CITIZENSHIP <b>CANADA</b>
POST OFFICE ADDRESS <b>3963 W. 24<sup>TH</sup> Avenue</b>		CITY <b>VANCOUVER</b>	STATE/COUNTRY ZIP CODE <b>CANADA V6S 1M1</b>
DATE <b>July 1, 1999</b>		SIGNATURE 	

- [X] Signature for additional joint inventor attached. Number of Pages 1.  
 [ ] Signature by Administrator(trix) or legal representative for deceased or incapacitated inventor. Number of Pages \_\_\_\_.  
 [ ] Signature for inventor who refuses to sign or cannot be reached by person authorized under 37 CFR § 1.47. Number of Pages \_\_\_\_.

OPPEDAHL & LARSON

FILE NO. NMED.P-001-2

NAME OF SECOND INVENTOR	LAST NAME BAILLIE	FIRST NAME DAVID	MIDDLE NAME L.
RESIDENCE & CITIZENSHIP	CITY OF RESIDENCE VANCOUVER	STATE OR COUNTRY OF RESIDENCE CANADA	COUNTRY OF CITIZENSHIP CANADA
POST OFFICE ADDRESS 29 North Kootenay Street		CITY VANCOUVER	STATE/COUNTRY ZIP CODE CANADA V5K 3P7
DATE July 1, 1999		SIGNATURE David L. Baillie	
NAME OF THIRD INVENTOR	LAST NAME	FIRST NAME	MIDDLE NAME
RESIDENCE & CITIZENSHIP	CITY OF RESIDENCE	STATE OR COUNTRY OF RESIDENCE	COUNTRY OF CITIZENSHIP
POST OFFICE ADDRESS		CITY	STATE/COUNTRY ZIP CODE
DATE		SIGNATURE	
NAME OF FOURTH INVENTOR	LAST NAME	FIRST NAME	MIDDLE NAME
RESIDENCE & CITIZENSHIP	CITY OF RESIDENCE	STATE OR COUNTRY OF RESIDENCE	COUNTRY OF CITIZENSHIP
POST OFFICE ADDRESS		CITY	STATE/COUNTRY ZIP CODE
DATE		SIGNATURE	
NAME OF FIFTH INVENTOR	LAST NAME	FIRST NAME	MIDDLE NAME
RESIDENCE & CITIZENSHIP	CITY OF RESIDENCE	STATE OR COUNTRY OF RESIDENCE	COUNTRY OF CITIZENSHIP
POST OFFICE ADDRESS		CITY	STATE/COUNTRY ZIP CODE
DATE		SIGNATURE	